

NUCLEIC ACID CONSTRUCTS AND METHODS FOR PRODUCING ALTERED SEED OIL COMPOSITIONS

CROSS-REFERENCE TO RELATED APPLICATIONS

This application is a continuation-in-part of U.S. Application No. 10/393,347, filed March 21,
5 2003, which application claims the benefit under 35 U.S.C. § 119(e) of U.S. Provisional Application Nos.
60/365,794 filed March 21, 2002, and 60/390,185 filed June 21, 2002, each of which is herein
incorporated by reference in its entirety.

INCORPORATION OF SEQUENCE LISTING

A paper copy of the Sequence Listing and a computer readable form of the sequence listing on
10 diskette, containing the file named "Omni2 AS FILED.txt", which is 60,690 bytes in size (measured in
MS-DOS), and which was recorded on September 25, 2003, are herein incorporated by reference.

FIELD OF THE INVENTION

The present invention is directed to recombinant nucleic acid molecules, constructs, and other
agents associated with the coordinate manipulation of multiple genes in the fatty acid synthesis pathway.
15 In particular, the agents of the present invention are associated with the simultaneous enhanced
expression of certain genes in the fatty acid synthesis pathway and suppressed expression of certain other
genes in the same pathway. The present invention is also directed to plants incorporating such agents,
and in particular to plants incorporating such constructs where the plants exhibit altered seed oil
compositions.

20

BACKGROUND

Plant oils are used in a variety of applications. Novel vegetable oil compositions and improved
approaches to obtain oil compositions, from biosynthetic or natural plant sources, are needed. Depending
upon the intended oil use, various different fatty acid compositions are desired. Plants, especially species
which synthesize large amounts of oils in seeds, are an important source of oils both for edible and
25 industrial uses. Seed oils are composed almost entirely of triacylglycerols in which fatty acids are
esterified to the three hydroxyl groups of glycerol.

Soybean oil typically contains about 16-20% saturated fatty acids: 13-16% palmitate and 3-4% stearate. See generally Gunstone *et al.*, *The Lipid Handbook*, Chapman & Hall, London (1994). Soybean oils have been modified by various breeding methods to create benefits for specific markets. However, a soybean oil that is broadly beneficial to major soybean oil users such as consumers of salad oil, cooking oil and frying oil, and industrial markets such as biodiesel and biolube markets, is not available. Prior soybean oils were either too expensive or lacked an important food quality property such as oxidative stability, good fried food flavor or saturated fat content, or an important biodiesel property such as appropriate nitric oxide emissions or cold tolerance or cold flow.

Higher plants synthesize fatty acids via a common metabolic pathway -- the fatty acid synthetase (FAS) pathway, which is located in the plastids. β -ketoacyl-ACP synthases are important rate-limiting enzymes in the FAS of plant cells and exist in several versions. β -ketoacyl-ACP synthase I catalyzes chain elongation to palmitoyl-ACP (C16:0), whereas β -ketoacyl-ACP synthase II catalyzes chain elongation to stearoyl-ACP (C18:0). β -ketoacyl-ACP synthase IV is a variant of β -ketoacyl-ACP synthase II, and can also catalyze chain elongation to 18:0-ACP. In soybean, the major products of FAS are 16:0-ACP and 18:0-ACP. The desaturation of 18:0-ACP to form 18:1-ACP is catalyzed by a plastid-localized soluble delta-9 desaturase (also referred to as "stearoyl-ACP desaturase"). See Voelker *et al.*, *52 Annu. Rev. Plant Physiol. Plant Mol. Biol.* 335-61 (2001).

The products of the plastidial FAS and delta-9 desaturase, 16:0-ACP, 18:0-ACP, and 18:1-ACP, are hydrolyzed by specific thioesterases (FAT). Plant thioesterases can be classified into two gene families based on sequence homology and substrate preference. The first family, FATA, includes long chain acyl-ACP thioesterases having activity primarily on 18:1-ACP. Enzymes of the second family, FATB, commonly utilize 16:0-ACP (palmitoyl-ACP), 18:0-ACP (stearoyl-ACP), and 18:1-ACP (oleoyl-ACP). Such thioesterases have an important role in determining chain length during de novo fatty acid biosynthesis in plants, and thus these enzymes are useful in the provision of various modifications of fatty acyl compositions, particularly with respect to the relative proportions of various fatty acyl groups that are present in seed storage oils.

The products of the FATA and FATB reactions, the free fatty acids, leave the plastids and are converted to their respective acyl-CoA esters. Acyl-CoAs are substrates for the lipid-biosynthesis pathway (Kennedy Pathway), which is located in the endoplasmic reticulum (ER). This pathway is

responsible for membrane lipid formation as well as the biosynthesis of triacylglycerols, which constitute the seed oil. In the ER there are additional membrane-bound desaturases, which can further desaturate 18:1 to polyunsaturated fatty acids. A delta-12 desaturase (FAD2) catalyzes the insertion of a double bond into 18:1, forming linoleic acid (18:2). A delta-15 desaturase (FAD3) catalyzes the insertion of a double bond into 18:2, forming linolenic acid (18:3).

Many complex biochemical pathways have now been manipulated genetically, usually by suppression or over-expression of single genes. Further exploitation of the potential for plant genetic manipulation will require the coordinate manipulation of multiple genes in a pathway. A number of approaches have been used to combine transgenes in one plant – including sexual crossing, retransformation, co-transformation, and the use of linked transgenes. A chimeric transgene with linked partial gene sequences can be used to coordinately suppress numerous plant endogenous genes. Constructs modeled on viral polyproteins can be used to simultaneously introduce multiple coding genes into plant cells. For a review, see Halpin *et al.*, *Plant Mol. Biol.* 47:295-310 (2001).

Thus, a desired plant phenotype may require the expression of one or more genes and the concurrent reduction of expression of another gene or genes. Thus, there exists a need to simultaneously over-express one or more genes and suppress, or down-regulate, the expression of a another gene or genes in plants using a single transgenic construct.

SUMMARY OF THE INVENTION

The present invention provides a nucleic acid molecule or molecules, which when introduced into a cell or organism are capable of suppressing, at least partially reducing, reducing, substantially reducing, or effectively eliminating the expression of at least one or more endogenous *FAD2*, *FAD3*, or *FATB* RNAs while at the same time coexpressing, simultaneously expressing, or coordinately producing one or more RNAs or proteins transcribed from or encoded by beta-ketoacyl-ACP synthase I, beta-ketoacyl-ACP synthase IV, delta-9 desaturase, or CP4 EPSPS. The present invention also provides plant cells and plants transformed with the same nucleic acid molecule or molecules, and seeds, oil, and other products produced from the transformed plants.

Also provided by the present invention is a recombinant nucleic acid molecule comprising a first set of DNA sequences that is capable, when expressed in a host cell, of suppressing the endogenous

expression of at least one, preferably two, genes selected from the group consisting of *FAD2*, *FAD3*, and *FATB* genes; and a second set of DNA sequences that is capable, when expressed in a host cell, of increasing the endogenous expression of at least one gene selected from the group consisting of a beta-ketoacyl-ACP synthase I gene, a beta-ketoacyl-ACP synthase IV gene, and a delta-9 desaturase gene.

5 Further provided by the present invention is a recombinant nucleic acid molecule comprising a first set of DNA sequences that is capable, when expressed in a host cell, of forming a dsRNA construct and suppressing the endogenous expression of at least one, preferably two, genes selected from the group consisting of *FAD2*, *FAD3*, and *FATB* genes; where the first set of DNA sequences comprises a first non-coding sequence that expresses a first RNA sequence that exhibits at least 90% identity to a non-coding
10 region of a *FAD2* gene, a first antisense sequence that expresses a first antisense RNA sequence capable of forming a double-stranded RNA molecule with the first RNA sequence, a second non-coding sequence that expresses a second RNA sequence that exhibits at least 90% identity to a non-coding region of a *FAD3* gene, and a second antisense sequence that expresses a second antisense RNA sequence capable of forming a double-stranded RNA molecule with the second RNA sequence; and a second set of DNA
15 sequences that is capable, when expressed in a host cell, of increasing the endogenous expression of at least one gene selected from the group consisting of a beta-ketoacyl-ACP synthase I gene, a beta-ketoacyl-ACP synthase IV gene, and a delta-9 desaturase gene.

The present invention provides methods of transforming plants with these recombinant nucleic acid molecules. The methods include a method of producing a transformed plant having seed with an
20 increased oleic acid content, reduced saturated fatty acid content, and reduced polyunsaturated fatty acid content, comprising (A) transforming a plant cell with a recombinant nucleic acid molecule which comprises a first set of DNA sequences that is capable, when expressed in a host cell, of suppressing the endogenous expression of at least one, preferably two, genes selected from the group consisting of *FAD2*, *FAD3*, and *FATB* genes, and a second set of DNA sequences that is capable, when expressed in a host
25 cell, of increasing the endogenous expression of at least one gene selected from the group consisting of a beta-ketoacyl-ACP synthase I gene, a beta-ketoacyl-ACP synthase IV gene, and a delta-9 desaturase gene; and (B) growing the transformed plant, where the transformed plant produces seed with an increased oleic acid content, reduced saturated fatty acid content, and reduced polyunsaturated fatty acid

content relative to seed from a plant having a similar genetic background but lacking the recombinant nucleic acid molecule.

Further provided are methods of transforming plant cells with the recombinant nucleic acid molecules. The methods include a method of altering the oil composition of a plant cell comprising (A) transforming a plant cell with a recombinant nucleic acid molecule which comprises a first set of DNA sequences that is capable, when expressed in a host cell, of suppressing the endogenous expression of at least one, preferably two, genes selected from the group consisting of *FAD2*, *FAD3*, and *FATB* genes, and a second set of DNA sequences that is capable, when expressed in a host cell, of increasing the endogenous expression of at least one gene selected from the group consisting of a beta-ketoacyl-ACP synthase I gene, a beta-ketoacyl-ACP synthase IV gene, and a delta-9 desaturase gene; and (B) growing the plant cell under conditions where transcription of the first set of DNA sequences and the second set of DNA sequences is initiated, where the oil composition is altered relative to a plant cell with a similar genetic background but lacking the recombinant nucleic acid molecule.

The present invention also provides a transformed plant comprising a recombinant nucleic acid molecule which comprises a first set of DNA sequences that is capable, when expressed in a host cell, of suppressing the endogenous expression of at least one, preferably two, genes selected from the group consisting of *FAD2*, *FAD3*, and *FATB* genes, and a second set of DNA sequences that is capable, when expressed in a host cell, of increasing the endogenous expression of at least one gene selected from the group consisting of a beta-ketoacyl-ACP synthase I gene, a beta-ketoacyl-ACP synthase IV gene, and a delta-9 desaturase gene. Further provided by the present invention is a transformed soybean plant bearing seed, where the seed exhibits an oil composition which comprises 55 to 80% by weight oleic acid, 10 to 40% by weight linoleic acid, 6% or less by weight linolenic acid, and 2 to 8% by weight saturated fatty acids, and feedstock, plant parts, and seed derived from the plant. In another embodiment, the present invention provides a transformed soybean plant bearing seed, where the seed exhibits an oil composition which comprises about 65-80% oleic acid, about 3-8% saturates, and about 10-20% polyunsaturates. Also included is feedstock, plant parts, and seed derived from such plant. In another embodiment, the present invention provides a transformed soybean plant bearing seed, where the seed exhibits an oil composition which comprises about 65-80% oleic acid, about 2-3.5% saturates, and about 10-25% polyunsaturates. Also included is feedstock, plant parts, and seed derived from such plant.

The present invention provides a soybean seed exhibiting an oil composition comprising 55 to 80% by weight oleic acid, 10 to 40% by weight linoleic acid, 6% or less by weight linolenic acid, and 2 to 8% by weight saturated fatty acids, and also provides a soybean seed exhibiting an oil composition comprising 65 to 80% by weight oleic acid, 10 to 30% by weight linoleic acid, 6% or less by weight linolenic acid, and 2 to 8% by weight of saturated fatty acids. In another embodiment, the present invention provides a soybean seed exhibiting an oil composition comprising about 65-80% oleic acid, about 3-8% saturates, and about 10-20% polyunsaturates. In another embodiment, the present invention provides a soybean seed exhibiting an oil composition which comprises about 65-80% oleic acid, about 2-3.5% saturates, and about 10-25% polyunsaturates.

Also provided by the present invention are soyfoods comprising an oil composition which comprises 69 to 73% by weight oleic acid, 21 to 24% by weight linoleic acid, 0.5 to 3% by weight linolenic acid, and 2-3% by weight of saturated fatty acids.

The crude soybean oil provided by the present invention exhibits an oil composition comprising 55 to 80% by weight oleic acid, 10 to 40% by weight linoleic acid, 6% or less by weight linolenic acid, and 2 to 8% by weight saturated fatty acids. Another crude soybean oil provided by the present invention exhibits an oil composition comprising 65 to 80% by weight oleic acid, 10 to 30% by weight linoleic acid, 6% or less by weight linolenic acid, and 2 to 8% by weight of saturated fatty acids. In another embodiment, the crude soybean oil provided by the present invention exhibits an oil composition comprising about 65-80% oleic acid, about 3-8% saturates, and about 10-20% polyunsaturates. In another embodiment, the crude soybean oil provided by the present invention exhibits an oil composition comprising about 65-80% oleic acid, about 2-3.5% saturates, and about 10-25% polyunsaturates.

BRIEF DESCRIPTION OF THE DRAWINGS

FIGS. 1-4 each depict exemplary nucleic acid molecule configurations.

FIGS. 5(a)-(d) and 6(a)-(c) each depict illustrative configurations of a first set of DNA sequences.

FIGS. 7- 20 each depict nucleic acid molecules of the present invention.

FIG. 21 depicts the construct pMON68537.

FIG. 22 depicts the construct pMON68539.

DETAILED DESCRIPTION OF THE INVENTION

Description of the Nucleic Acid Sequences

- SEQ ID NO: 1 is a nucleic acid sequence of a *FAD2-1A* intron 1.
- SEQ ID NO: 2 is a nucleic acid sequence of a *FAD2-1B* intron 1.
- 5 SEQ ID NO: 3 is a nucleic acid sequence of a *FAD2-1B* promoter.
- SEQ ID NO: 4 is a nucleic acid sequence of a *FAD2-1A* genomic clone.
- SEQ ID NOS: 5 & 6 are nucleic acid sequences of a *FAD2-1A* 3' UTR and 5'UTR, respectively.
- SEQ ID NOS: 7-13 are nucleic acid sequences of *FAD3-1A* introns 1, 2, 3A, 4, 5, 3B, and 3C,
respectively.
- 10 SEQ ID NO: 14 is a nucleic acid sequence of a *FAD3-1C* intron 4.
- SEQ ID NO: 15 is a nucleic acid sequence of a partial *FAD3-1A* genomic clone.
- SEQ ID NOS: 16 & 17 are nucleic acid sequences of a *FAD3-1A* 3'UTR and 5'UTR, respectively.
- SEQ ID NO: 18 is a nucleic acid sequence of a partial *FAD3-1B* genomic clone.
- SEQ ID NOS: 19-25 are nucleic acid sequences of *FAD3-1B* introns 1, 2, 3A, 3B, 3C, 4, and 5,
15 respectively.
- SEQ ID NOS: 26 & 27 are nucleic acid sequences of a *FAD3-1B* 3'UTR and 5'UTR, respectively.
- SEQ ID NO: 28 is a nucleic acid sequence of a *FATB-1* genomic clone.
- SEQ ID NO: 29-35 are nucleic acid sequences of *FATB-1* introns I, II, III, IV, V, VI, and VII,
respectively.
- 20 SEQ ID NOS: 36 & 37 are nucleic acid sequences of a *FATB-1* 3'UTR and 5'UTR, respectively.
- SEQ ID NO: 38 is a nucleic acid sequence of a *Cuphea pulcherrima KAS I* gene.
- SEQ ID NO: 39 is a nucleic acid sequence of a *Cuphea pulcherrima KAS IV* gene.
- SEQ ID NOS: 40 & 41 are nucleic acid sequences of *Ricinus communis* and *Simmondsia
chinensis* delta-9 desaturase genes, respectively.
- 25 SEQ ID NO: 42 is a nucleic acid sequence of a *FATB-2* cDNA.
- SEQ ID NO: 43 is a nucleic acid sequence of a *FATB-2* genomic clone.
- SEQ ID NOS: 44-47 are nucleic acid sequences of *FATB-2* introns I, II, III, and IV respectively.
- SEQ ID NOS: 48-60 are nucleic acid sequences of PCR primers.

Definitions

“ACP” refers to an acyl carrier protein moiety. “Altered seed oil composition” refers to a seed oil composition from a transgenic or transformed plant of the invention which has altered or modified levels of the fatty acids therein, relative to a seed oil from a plant having a similar genetic background but that has not been transformed. “Antisense suppression” refers to gene-specific silencing that is induced by the introduction of an antisense RNA molecule.

“Coexpression of more than one agent such as an mRNA or protein” refers to the simultaneous expression of an agent in overlapping time frames and in the same cell or tissue as another agent.

“Coordinated expression of more than one agent” refers to the coexpression of more than one agent when the production of transcripts and proteins from such agents is carried out utilizing a shared or identical promoter. “Complement” of a nucleic acid sequence refers to the complement of the sequence along its complete length.

“Cosuppression” is the reduction in expression levels, usually at the level of RNA, of a particular endogenous gene or gene family by the expression of a homologous sense construct that is capable of transcribing mRNA of the same strandedness as the transcript of the endogenous gene. Napoli *et al.*, *Plant Cell* 2:279-289 (1990); van der Krol *et al.*, *Plant Cell* 2:291-299 (1990). “Crude soybean oil” refers to soybean oil that has been extracted from soybean seeds, but has not been refined, processed, or blended, although it may be degummed.

When referring to proteins and nucleic acids herein, “derived” refers to either directly (for example, by looking at the sequence of a known protein or nucleic acid and preparing a protein or nucleic acid having a sequence similar, at least in part, to the sequence of the known protein or nucleic acid) or indirectly (for example, by obtaining a protein or nucleic acid from an organism which is related to a known protein or nucleic acid) obtaining a protein or nucleic acid from a known protein or nucleic acid. Other methods of “deriving” a protein or nucleic acid from a known protein or nucleic acid are known to one of skill in the art.

“dsRNA”, “dsRNAi” and “RNAi” all refer to gene-specific silencing that is induced by the introduction of a construct capable of forming a double-stranded RNA molecule. A “dsRNA molecule” and an “RNAi molecule” both refer to a double-stranded RNA molecule capable, when introduced into a

cell or organism, of at least partially reducing the level of an mRNA species present in a cell or a cell of an organism.

“Exon” refers to the normal sense of the term as meaning a segment of nucleic acid molecules, usually DNA, that encodes part of or all of an expressed protein.

5 “Fatty acid” refers to free fatty acids and fatty acyl groups.

“Gene” refers to a nucleic acid sequence that encompasses a 5’ promoter region associated with the expression of the gene product, any intron and exon regions and 3’ or 5’ untranslated regions associated with the expression of the gene product. “Gene silencing” refers to the suppression of gene expression or down-regulation of gene expression.

10 A “gene family” is two or more genes in an organism which encode proteins that exhibit similar functional attributes, and a “gene family member” is any gene of the gene family found within the genetic material of the plant, e.g., a “*FAD2* gene family member” is any *FAD2* gene found within the genetic material of the plant. An example of two members of a gene family are *FAD2-1* and *FAD2-2*. A gene family can be additionally classified by the similarity of the nucleic acid sequences. Preferably, a gene
15 family member exhibits at least 60%, more preferably at least 70%, more preferably at least 80% nucleic acid sequence identity in the coding sequence portion of the gene.

“Heterologous” means not naturally occurring together. A “high oleic soybean seed” is a seed with oil having greater than 75% oleic acid present in the oil composition of the seed.

A nucleic acid molecule is said to be “introduced” if it is inserted into a cell or organism as a
20 result of human manipulation, no matter how indirect. Examples of introduced nucleic acid molecules include, but are not limited to, nucleic acids that have been introduced into cells via transformation, transfection, injection, and projection, and those that have been introduced into an organism via methods including, but not limited to, conjugation, endocytosis, and phagocytosis.

“Intron” refers to the normal sense of the term as meaning a segment of nucleic acid molecules,
25 usually DNA, that does not encode part of or all of an expressed protein, and which, in endogenous conditions, is transcribed into RNA molecules, but which is spliced out of the endogenous RNA before the RNA is translated into a protein. An “intron dsRNA molecule” and an “intron RNAi molecule” both refer to a double-stranded RNA molecule capable, when introduced into a cell or organism, of at least partially reducing the level of an mRNA species present in a cell or a cell of an organism where the

double-stranded RNA molecule exhibits sufficient identity to an intron of a gene present in the cell or organism to reduce the level of an mRNA containing that intron sequence.

A "low saturate" oil composition contains between 3.6 and 8 percent saturated fatty acids.

5 A "mid-oleic soybean seed" is a seed having between 50% and 85% oleic acid present in the oil composition of the seed.

The term "non-coding" refers to sequences of nucleic acid molecules that do not encode part or all of an expressed protein. Non-coding sequences include but are not limited to introns, promoter regions, 3' untranslated regions (3'UTRs), and 5' untranslated regions (5'UTRs).

10 A promoter that is "operably linked" to one or more nucleic acid sequences is capable of driving expression of one or more nucleic acid sequences, including multiple coding or non-coding nucleic acid sequences arranged in a polycistronic configuration.

15 "Physically linked" nucleic acid sequences are nucleic acid sequences that are found on a single nucleic acid molecule. A "plant" includes reference to whole plants, plant organs (e.g., leaves, stems, roots, etc.), seeds, and plant cells and progeny of the same. The term "plant cell" includes, without limitation, seed suspension cultures, embryos, meristematic regions, callus tissue, leaves, roots, shoots, gametophytes, sporophytes, pollen, and microspores. "Plant promoters," include, without limitation, plant viral promoters, promoters derived from plants, and synthetic promoters capable of functioning in a plant cell to promote the expression of an mRNA.

20 A "polycistronic gene" or "polycistronic mRNA" is any gene or mRNA that contains transcribed nucleic acid sequences which correspond to nucleic acid sequences of more than one gene targeted for expression. It is understood that such polycistronic genes or mRNAs may contain sequences that correspond to introns, 5'UTRs, 3'UTRs, or combinations thereof, and that a recombinant polycistronic gene or mRNA might, for example without limitation, contain sequences that correspond to one or more UTRs from one gene and one or more introns from a second gene.

25 A "seed-specific promoter" refers to a promoter that is active preferentially or exclusively in a seed. "Preferential activity" refers to promoter activity that is substantially greater in the seed than in other tissues, organs or organelles of the plant. "Seed-specific" includes without limitation activity in the aleurone layer, endosperm, and/or embryo of the seed.

“Sense intron suppression” refers to gene silencing that is induced by the introduction of a sense intron or fragment thereof. Sense intron suppression is described, for example by Fillatti in PCT WO 01/14538 A2. “Simultaneous expression” of more than one agent such as an mRNA or protein refers to the expression of an agent at the same time as another agent. Such expression may only overlap in part and may also occur in different tissue or at different levels.

“Total oil level” refers to the total aggregate amount of fatty acid without regard to the type of fatty acid. “Transgene” refers to a nucleic acid sequence associated with the expression of a gene introduced into an organism. A transgene includes, but is not limited to, a gene endogenous or a gene not naturally occurring in the organism. A “transgenic plant” is any plant that stably incorporates a transgene in a manner that facilitates transmission of that transgene from a plant by any sexual or asexual method.

A “zero saturate” oil composition contains less than 3.6 percent saturated fatty acids.

When referring to proteins and nucleic acids herein, the use of plain capitals, e.g., “FAD2”, indicates a reference to an enzyme, protein, polypeptide, or peptide, and the use of italicized capitals, e.g., “*FAD2*”, is used to refer to nucleic acids, including without limitation genes, cDNAs, and mRNAs. A cell or organism can have a family of more than one gene encoding a particular enzyme, and the capital letter that follows the gene terminology (A, B, C) is used to designate the family member, i.e., *FAD2-1A* is a different gene family member from *FAD2-1B*.

As used herein, any range set forth is inclusive of the end points of the range unless otherwise stated.

A. Agents

The agents of the invention will preferably be “biologically active” with respect to either a structural attribute, such as the capacity of a nucleic acid molecule to hybridize to another nucleic acid molecule, or the ability of a protein to be bound by an antibody (or to compete with another molecule for such binding). Alternatively, such an attribute may be catalytic and thus involve the capacity of the agent to mediate a chemical reaction or response. The agents will preferably be “substantially purified.” The term “substantially purified,” as used herein, refers to a molecule separated from substantially all other molecules normally associated with it in its native environmental conditions. More preferably a substantially purified molecule is the predominant species present in a preparation. A substantially purified molecule may be greater than 60% free, greater than 75% free, preferably greater than 90% free,

and most preferably greater than 95% free from the other molecules (exclusive of solvent) present in the natural mixture. The term "substantially purified" is not intended to encompass molecules present in their native environmental conditions.

5 The agents of the invention may also be recombinant. As used herein, the term "recombinant" means any agent (*e.g.*, including but limited to DNA, peptide), that is, or results, however indirectly, from human manipulation of a nucleic acid molecule. It is also understood that the agents of the invention may be labeled with reagents that facilitate detection of the agent, *e.g.*, fluorescent labels, chemical labels, and/or modified bases.

10 Agents of the invention include nucleic acid molecules that comprise a DNA sequence which is at least 50%, 60%, or 70% identical over their entire length to a plant coding region or non-coding region, or to a nucleic acid sequence that is complementary to a plant coding or non-coding region. More preferable are DNA sequences that are, over their entire length, at least 80% identical; at least 85% identical; at least 90% identical; at least 95% identical; at least 97% identical; at least 98% identical; at least 99% identical; or 100% identical to a plant coding region or non-coding region, or to a nucleic acid sequence that is
15 complementary to a plant coding or non-coding region.

"Identity," as is well understood in the art, is a relationship between two or more polypeptide sequences or two or more nucleic acid molecule sequences, as determined by comparing the sequences. In the art, "identity" also means the degree of sequence relatedness between polypeptide or nucleic acid molecule sequences, as determined by the match between strings of such sequences. "Identity" can be
20 readily calculated by known methods including, but not limited to, those described in *Computational Molecular Biology*, Lesk, ed., Oxford University Press, New York 1988; *Biocomputing: Informatics and Genome Projects*, Smith, ed., Academic Press, New York 1993; *Computer Analysis of Sequence Data, Part I*, Griffin and Griffin, eds., Humana Press, New Jersey 1994; *Sequence Analysis in Molecular Biology*, von Heinje, Academic Press 1987; *Sequence Analysis Primer*, Gribskov and Devereux, eds.,
25 Stockton Press, New York 1991; and Carillo and Lipman, *SIAM J. Applied Math*, 48:1073 1988.

Methods to determine identity are designed to give the largest match between the sequences tested. Moreover, methods to determine identity are codified in publicly available programs. Computer programs which can be used to determine identity between two sequences include, but are not limited to, GCG; a suite of five BLAST programs, three designed for nucleotide sequences queries (BLASTN,

BLASTX, and TBLASTX) and two designed for protein sequence queries (BLASTP and TBLASTN). The BLASTX program is publicly available from NCBI and other sources, *e.g.*, *BLAST Manual*, Altschul *et al.*, NCBI NLM NIH, Bethesda, MD 20894; Altschul *et al.*, *J. Mol. Biol.* 215:403-410 (1990). The well-known Smith Waterman algorithm can also be used to determine identity.

5 Parameters for polypeptide sequence comparison typically include the following: Algorithm: Needleman and Wunsch, *J. Mol. Biol.* 48:443-453 (1970); Comparison matrix: BLOSSUM62 from Hentikoff and Hentikoff, *Proc. Natl. Acad. Sci. USA* 89:10915-10919 (1992); Gap Penalty: 12; Gap Length Penalty: 4. A program that can be used with these parameters is publicly available as the "gap" program from Genetics Computer Group ("GCG"), Madison, Wisconsin. The above parameters along
10 with no penalty for end gap are the default parameters for peptide comparisons.

 Parameters for nucleic acid molecule sequence comparison include the following: Algorithm: Needleman and Wunsch, *J. Mol. Bio.* 48:443-453 (1970); Comparison matrix: matches - +10; mismatches = 0; Gap Penalty: 50; Gap Length Penalty: 3. As used herein, "% identity" is determined using the above parameters as the default parameters for nucleic acid molecule sequence comparisons and the "gap"
15 program from GCG, version 10.2.

 Subsets of the nucleic acid sequences of the present invention include fragment nucleic acid molecules. "Fragment nucleic acid molecule" refers to a piece of a larger nucleic acid molecule, which may consist of significant portion(s) of, or indeed most of, the larger nucleic acid molecule, or which may comprise a smaller oligonucleotide having from about 15 to about 400 contiguous nucleotides and more.
20 preferably, about 15 to about 45 contiguous nucleotides, about 20 to about 45 contiguous nucleotides, about 15 to about 30 contiguous nucleotides, about 21 to about 30 contiguous nucleotides, about 21 to about 25 contiguous nucleotides, about 21 to about 24 contiguous nucleotides, about 19 to about 25 contiguous nucleotides, or about 21 contiguous nucleotides. Fragment nucleic acid molecules may consist of significant portion(s) of, or indeed most of, a plant coding or non-coding region, or
25 alternatively may comprise smaller oligonucleotides. In a preferred embodiment, a fragment shows 100% identity to the plant coding or non-coding region. In another preferred embodiment, a fragment comprises a portion of a larger nucleic acid sequence. In another aspect, a fragment nucleic acid molecule has a nucleic acid sequence that has at least 15, 25, 50, or 100 contiguous nucleotides of a nucleic acid molecule of the present invention. In a preferred embodiment, a nucleic acid molecule has a

nucleic acid sequence that has at least 15, 25, 50, or 100 contiguous nucleotides of a plant coding or non-coding region.

In another aspect of the present invention, the DNA sequence of the nucleic acid molecules of the present invention can comprise sequences that differ from those encoding a polypeptide or fragment of the protein due to conservative amino acid changes in the polypeptide; the nucleic acid sequences coding for the polypeptide can therefore have sequence differences corresponding to the conservative changes.

In a further aspect of the present invention, one or more of the nucleic acid molecules of the present invention differ in nucleic acid sequence from those for which a specific sequence is provided herein because one or more codons have been replaced with a codon that encodes a conservative substitution of the amino acid originally encoded.

Agents of the invention also include nucleic acid molecules that encode at least about a contiguous 10 amino acid region of a polypeptide of the present invention, more preferably at least about a contiguous 25, 40, 50, 100, or 125 amino acid region of a polypeptide of the present invention. Due to the degeneracy of the genetic code, different nucleotide codons may be used to code for a particular amino acid. A host cell often displays a preferred pattern of codon usage. Structural nucleic acid sequences are preferably constructed to utilize the codon usage pattern of the particular host cell. This generally enhances the expression of the structural nucleic acid sequence in a transformed host cell. Any of the above-described nucleic acid and amino acid sequences may be modified to reflect the preferred codon usage of a host cell or organism in which they are contained. Therefore, a contiguous 10 amino acid region of a polypeptide of the present invention could be encoded by numerous different nucleic acid sequences. Modification of a structural nucleic acid sequence for optimal codon usage in plants is described in U.S. Patent No. 5,689,052.

Agents of the invention include nucleic acid molecules. For example, without limitation, in an aspect of the present invention, the nucleic acid molecule of the present invention comprises an intron sequence of SEQ ID NO: 19, 20, 21, 22, 23, 25, 32, 33, 34, 35, 44, 45, 46, or 47 or fragments thereof or complements thereof. In another aspect of the invention, the nucleic acid molecule comprises a nucleic acid sequence, which when introduced into a cell or organism, is capable of suppressing the production of an RNA or protein while simultaneously expressing, coexpressing or coordinately expressing another RNA or protein. In an aspect of the invention, the nucleic acid molecule comprises a nucleic acid

sequence, which when introduced into a cell or organism is capable of suppressing, at least partially reducing, reducing, substantially reducing, or effectively eliminating the expression of endogenous *FAD2*, *FAD3*, and/or *FATB* RNA while at the same time coexpressing, simultaneously expressing, or coordinately expressing a beta-ketoacyl-ACP synthase I, beta-ketoacyl-ACP synthase IV, delta-9 desaturase, and/or CP4 EPSPS RNA or protein.

By decreasing the amount of *FAD2* and/or *FAD3* available in a plant cell, a decreased percentage of polyunsaturated fatty acids such as linoleate (C18:2) and linolenate (C18:3) may be provided. Modifications in the pool of fatty acids available for incorporation into triacylglycerols may likewise affect the composition of oils in the plant cell. Thus, a decrease in expression of *FAD2* and/or *FAD3* may result in an increased proportion of mono-unsaturated fatty acids such as oleate (C18:1). When the amount of *FATB* is decreased in a plant cell, a decreased amount of saturated fatty acids such as palmitate and stearate may be provided. Thus, a decrease in expression of *FATB* may result in an increased proportion of unsaturated fatty acids such as oleate (18:1). The simultaneous suppression of *FAD2*, *FAD3*, and *FATB* expression thereby results in driving the FAS pathway toward an overall increase in mono-unsaturated fatty acids of 18-carbon length, such as oleate (C18:1). See U.S. Patent No. 5,955,650.

By increasing the amount of beta-ketoacyl-ACP synthase I (KAS I) and/or beta-ketoacyl-ACP synthase IV (KAS IV) available in a plant cell, a decreased percentage of 16:0-ACP may be provided, leading to an increased percentage of 18:0-ACP. A greater amount of 18:0-ACP in combination with the simultaneous suppression of one or more of *FAD2*, *FAD3*, and *FATB*, thereby helps drive the oil composition toward an overall increase in oleate (C18:1). By increasing the amount of delta-9 desaturase available in a plant cell, an increased percentage of unsaturated fatty acids may be provided, resulting in an overall lowering of stearate and total saturates.

These combinations of increased and decreased enzyme expression may be manipulated to produce fatty acid compositions, including oils, having an increased oleate level, decreased linoleate, linolenate, stearate, and/or palmitate levels, and a decreased overall level of saturates. Enhancement of gene expression in plants may occur through the introduction of extra copies of coding sequences of the genes into the plant cell or, preferably, the incorporation of extra copies of coding sequences of the gene into the plant genome. Over-expression may also occur through increasing the activities of the regulatory mechanisms that regulate the expression of genes, *i.e.*, up-regulation of the gene expression.

Production of CP4 EPSPS in a plant cell provides the plant cell with resistance or tolerance to glyphosate, thereby providing a convenient method for identification of successful transformants via glyphosate-tolerant selection.

5 Suppression of gene expression in plants, also known as gene silencing, occurs at both the transcriptional level and post-transcriptional level. There are various methods for the suppression of expression of endogenous sequences in a host cell, including, but not limited to, antisense suppression, co-suppression, ribozymes, combinations of sense and antisense (double-stranded RNAi), promoter silencing, and DNA binding proteins such as zinc finger proteins. (See, e.g., WO 98/53083, WO 01/14538, and U.S. Patent 5,759,829 (Shewmaker.)). Certain of these mechanisms are associated with
10 nucleic acid homology at the DNA or RNA level. In plants, double-stranded RNA molecules can induce sequence-specific silencing. Gene silencing is often referred to as double stranded RNA ("dsRNAi") in plants, as RNA interference or RNAi in *Caenorhabditis elegans* and in animals, and as quelling in fungi.

 In a preferred embodiment, the nucleic acid molecule of the present invention comprises (a) a first set of DNA sequences, each of which exhibits sufficient homology to one or more coding or non-coding sequences of a plant gene such that when it is expressed, it is capable of effectively eliminating, substantially reducing, or at least partially reducing the level of an mRNA transcript or protein encoded by the gene from which the coding or non-coding sequence was derived, or any gene which has homology
15 to the target non-coding sequence, and (b) a second set of DNA sequences, each of which exhibits sufficient homology to a plant gene so that when it is expressed, it is capable of at least partially
20 enhancing, increasing, enhancing, or substantially enhancing the level of an mRNA transcript or protein encoded by the gene.

 As used herein, "a reduction" of the level or amount of an agent such as a protein or mRNA means that the level or amount is reduced relative to a cell or organism lacking a DNA sequence capable of reducing the agent. For example, "at least a partial reduction" refers to a reduction of at least 25%, "a
25 substantial reduction" refers to a reduction of at least 75%, and "an effective elimination" refers to a reduction of greater than 95%, all of which reductions in the level or amount of the agent are relative to a cell or organism lacking a DNA sequence capable of reducing the agent.

 As used herein, "an enhanced" or "increased" level or amount of an agent such as a protein or mRNA means that the level or amount is higher than the level or amount of agent present in a cell, tissue

or plant with a similar genetic background but lacking an introduced nucleic acid molecule encoding the protein or mRNA. For example, an “at least partially enhanced” level refers to an increase of at least 25%, and a “substantially enhanced” level refers to an increase of at least 100%, all of which increases in the level or amount of an agent are relative to the level or amount of agent that is present in a cell, tissue or plant with a similar genetic background but lacking an introduced nucleic acid molecule encoding the protein or mRNA.

When levels of an agent are compared, such a comparison is preferably carried out between organisms with a similar genetic background. Preferably, a similar genetic background is a background where the organisms being compared share 50% or greater, more preferably 75% or greater, and, even more preferably 90% or greater sequence identity of nuclear genetic material. In another preferred aspect, a similar genetic background is a background where the organisms being compared are plants, and the plants are isogenic except for any genetic material originally introduced using plant transformation techniques. Measurement of the level or amount of an agent may be carried out by any suitable method, non-limiting examples of which include comparison of mRNA transcript levels, protein or peptide levels, and/or phenotype, especially oil content. As used herein, mRNA transcripts include processed and non-processed mRNA transcripts, and proteins or peptides include proteins or peptides with or without any post-translational modification.

The DNA sequences of the first set of DNA sequences may be coding sequences, intron sequences, 3'UTR sequences, 5'UTR sequences, promoter sequences, other non-coding sequences, or any combination of the foregoing. The first set of DNA sequences encodes one or more sequences which, when expressed, are capable of selectively reducing either or both the protein and the transcript encoded by a gene selected from the group consisting of *FAD2*, *FAD3*, and *FATB*. In a preferred embodiment, the first set of DNA sequences is capable of expressing antisense RNA, in which the individual antisense sequences may be linked in one transcript, or may be in unlinked individual transcripts. In a further preferred embodiment, the first set of DNA sequences are physically linked sequences which are capable of expressing a single dsRNA molecule. In a different preferred embodiment, the first set of DNA sequences is capable of expressing sense cosuppression RNA, in which the individual sense sequences may be linked in one transcript, or may be in unlinked individual transcripts. Exemplary embodiments of the first set of DNA sequences are described in Part B of the Detailed Description, and in the Examples.

The second set of DNA sequences encodes one or more sequences which, when expressed, are capable of increasing one or both of the protein and transcript encoded by a gene selected from the group consisting of beta-ketoacyl-ACP synthase I (*KAS I*), beta-ketoacyl-ACP synthase IV (*KAS IV*), delta-9 desaturase, and CP4 EPSPS. The DNA sequences of the second set of DNA sequences may be physically
5 linked sequences. Exemplary embodiments of the second set of DNA sequences are described below in Parts C and D of the Detailed Description.

Thus, the present invention provides methods for altering the composition of fatty acids and compounds containing such fatty acids, such as oils, waxes, and fats. The present invention also provides methods for the production of particular fatty acids in host cell plants. Such methods employ the use of
10 the expression cassettes described herein for the modification of the host plant cell's FAS pathway.

B. First Set of DNA Sequences

In an aspect of the present invention, a nucleic acid molecule comprises a first set of DNA sequences, which when introduced into a cell or organism, expresses one or more sequences capable of effectively eliminating, substantially reducing, or at least partially reducing the levels of mRNA
15 transcripts or proteins encoded by one or more genes. Preferred aspects include as a target an endogenous gene, a plant gene, and a non-viral gene. In an aspect of the present invention, a gene is a *FAD2*, *FAD3*, or *FATB* gene.

In an aspect, a nucleic acid molecule of the present invention comprises a DNA sequence which exhibits sufficient homology to one or more coding or non-coding sequences from a plant gene, which
20 when introduced into a plant cell or plant and expressed, is capable of effectively eliminating, substantially reducing, or at least partially reducing the level of an mRNA transcript or protein encoded by the gene from which the coding or non-coding sequence(s) was derived. The DNA sequences of the first set of DNA sequences encode RNA sequences or RNA fragments which exhibit at least 90%, preferably at least 95%, more preferably at least 98%, most preferably at least 100% identity to a coding
25 or non-coding region derived from the gene which is to be suppressed. Such percent identity may be to a nucleic acid fragment.

Preferably, the non-coding sequence is a 3' UTR, 5'UTR, or a plant intron from a plant gene. More preferably, the non-coding sequence is a promoter sequence, 3' UTR, 5'UTR, or a plant intron from a plant gene. The intron may be located between exons, or located within a 5' or 3' UTR of a plant gene.

The sequence(s) of the first set of DNA sequences may be designed to express a dsRNA construct, a sense suppression RNA construct, or an antisense RNA construct or any other suppression construct in order to achieve the desired effect when introduced into a plant cell or plant. Such DNA sequence(s) may be fragment nucleic acid molecules. In a preferred aspect, a dsRNA construct contains
5 exon sequences, but the exon sequences do not correspond to a sufficient part of a plant exon to be capable of effectively eliminating, substantially reducing, or at least partially reducing the level of an mRNA transcript or protein encoded by the gene from which the exon was derived.

A plant intron can be any plant intron from a gene, whether endogenous or introduced. Nucleic acid sequences of such introns can be derived from a multitude of sources, including, without limitation,
10 databases such as EMBL and Genbank which may be found on the Internet at ebi.ac.uk/swisprot/; expasy.ch/; embl-heidelberg.de/; and ncbi.nlm.nih.gov. Nucleic acid sequences of such introns can also be derived, without limitation, from sources such as the GENSCAN program which may be found on the Internet at genes.mit.edu/GENSCAN.html.

Additional introns may also be obtained by methods which include, without limitation, screening
15 a genomic library with a probe of either known exon or intron sequences, comparing genomic sequence with its corresponding cDNA sequence, or cloning an intron such as a soybean intron by alignment to an intron from another organism, such as, for example, *Arabidopsis*. In addition, other nucleic acid sequences of introns will be apparent to one of ordinary skill in the art. The above-described methods may also be used to derive and obtain other non-coding sequences, including but not limited to, promoter
20 sequences, 3'UTR sequences, and 5'UTR sequences.

A "*FAD2*", " $\Delta 12$ desaturase" or "omega-6 desaturase" gene encodes an enzyme (*FAD2*) capable of catalyzing the insertion of a double bond into a fatty acyl moiety at the twelfth position counted from the carboxyl terminus. The term "*FAD2-1*" is used to refer to a *FAD2* gene that is naturally expressed in a specific manner in seed tissue, and the term "*FAD2-2*" is used to refer a *FAD2* gene that is (a)
25 different gene from a *FAD2-1* gene and (b) is naturally expressed in multiple tissues, including the seed. Representative *FAD2* sequences include, without limitation, those set forth in U.S. Patent Application No. 10/176,149 filed on June 21, 2002, and in SEQ ID NOs: 1-6.

A "*FAD3*", " $\Delta 15$ desaturase" or "omega-3 desaturase" gene encodes an enzyme (*FAD3*) capable of catalyzing the insertion of a double bond into a fatty acyl moiety at the fifteenth position counted from

the carboxyl terminus. The term "*FAD3-1*" is used to refer a *FAD3* gene family member that is naturally expressed in multiple tissues, including the seed. Representative *FAD3* sequences include, without limitation, those set forth in U.S. Patent Application No. 10/176,149 filed on June 21, 2002, and in SEQ ID NOs: 7-27.

5 A "*FATB*" or "palmitoyl-ACP thioesterase" gene encodes an enzyme (*FATB*) capable of catalyzing the hydrolytic cleavage of the carbon-sulfur thioester bond in the panthothene prosthetic group of palmitoyl-ACP as its preferred reaction. Hydrolysis of other fatty acid-ACP thioesters may also be catalyzed by this enzyme. Representative *FATB-1* sequences include, without limitation, those set forth in U.S. Provisional Application No. 60/390,185 filed on June 21, 2002; U.S. Patent Nos. 5,955,329;
10 5,723,761; 5,955,650; and 6,331,664; and SEQ ID NOs: 28-37. Representative *FATB-2* sequences include, without limitation, those set forth in SEQ ID NOs: 42-47.

C. Second Set of DNA Sequences

In an aspect of the present invention, a nucleic acid molecule comprises a second set of DNA sequences, which when introduced into a cell or organism, is capable of partially enhancing, increasing,
15 enhancing, or substantially enhancing the levels of mRNA transcripts or proteins encoded by one or more genes. In an aspect of the present invention, a gene is an endogenous gene. In an aspect of the present invention, a gene is a plant gene. In another aspect of the present invention, a gene is a truncated gene where the truncated gene is capable of catalyzing the reaction catalyzed by the full gene. In an aspect of the present invention, a gene is a beta-ketoacyl-ACP synthase I, beta-ketoacyl-ACP synthase IV, delta-9
20 desaturase, or CP4 *EPSPS* gene.

A gene of the present invention can be any gene, whether endogenous or introduced. Nucleic acid sequences of such genes can be derived from a multitude of sources, including, without limitation, databases such as EMBL and Genbank which may be found on the Internet at ebi.ac.uk/swisprot/;
expasy.ch/; embl-heidelberg.de/; and ncbi.nlm.nih.gov. Nucleic acid sequences of such genes can also be
25 derived, without limitation, from sources such as the GENSCAN program which may be found on the Internet at genes.mit.edu/GENSCAN.html.

Additional genes may also be obtained by methods which include, without limitation, screening a genomic library or a cDNA library with a probe of either known gene sequences, cloning a gene by alignment to a gene or probe from another organism, such as, for example, *Arabidopsis*. In addition,

other nucleic acid sequences of genes will be apparent to one of ordinary skill in the art. Additional genes may, for example without limitation, be amplified by polymerase chain reaction (PCR) and used in an embodiment of the present invention. In addition, other nucleic acid sequences of genes will be apparent to one of ordinary skill in the art.

5 Automated nucleic acid synthesizers may be employed for this purpose, and to make a nucleic acid molecule that has a sequence also found in a cell or organism. In lieu of such synthesis, nucleic acid molecules may be used to define a pair of primers that can be used with the PCR to amplify and obtain any desired nucleic acid molecule or fragment of a first gene.

10 A “*KAS I*” or “beta-ketoacyl-ACP synthase I” gene encodes an enzyme (KAS I) capable of catalyzing the elongation of a fatty acyl moiety up to palmitoyl-ACP (C16:0). Representative *KAS I* sequences include, without limitation, those set forth in U.S. Patent No. 5,475,099 and PCT Publication WO 94/10189, and in SEQ ID NO: 38.

15 A “*KAS IV*” or “beta-ketoacyl-ACP synthase IV” gene encodes an enzyme (KAS IV) capable of catalyzing the condensation of medium chain acyl-ACPs and enhancing the production of 18:0-ACP. Representative *KAS IV* sequences include, without limitation, those set forth in PCT Publication WO 98/46776, and in SEQ ID NO: 39.

20 A “delta-9 desaturase” or “stearoyl-ACP desaturase” or “omega-9 desaturase” gene encodes an enzyme capable of catalyzing the insertion of a double bond into a fatty acyl moiety at the ninth position counted from the carboxyl terminus. A preferred delta-9 desaturase of the present invention is a plant or cyanobacterial delta-9 desaturase, and more preferably a delta-9 desaturase that is also found in an organism selected from the group consisting of *Carthamus tinctorius*, *Ricinus communis*, *Simmonsia chinensis*, and *Brassica campestris*. Representative delta-9 desaturase sequences include, without limitation, those set forth in U.S. Patent No. 5,723,595, and SEQ ID NOs: 40-41.

25 A “CP4 *EPSPS*” or “CP4 5-enolpyruvylshikimate-3-phosphate synthase” gene encodes an enzyme (CP4 *EPSPS*) capable of conferring a substantial degree of glyphosate resistance upon the plant cell and plants generated therefrom. The CP4 *EPSPS* sequence may be a CP4 *EPSPS* sequence derived from *Agrobacterium tumefaciens* sp. CP4 or a variant or synthetic form thereof, as described in U.S. Patent No. 5,633,435. Representative CP4 *EPSPS* sequences include, without limitation, those set forth in U.S. Patent Nos. 5,627,061 and 5,633,435.

D. Recombinant Vectors and Constructs

One or more of the nucleic acid constructs of the invention may be used in plant transformation or transfection. The levels of products such as transcripts or proteins may be increased or decreased throughout an organism such as a plant or localized in one or more specific organs or tissues of the organism. For example the levels of products may be increased or decreased in one or more of the tissues and organs of a plant including without limitation: roots, tubers, stems, leaves, stalks, fruit, berries, nuts, bark, pods, seeds and flowers. A preferred organ is a seed. For example, exogenous genetic material may be transferred into a plant cell and the plant cell regenerated into a whole, fertile or sterile plant or plant part.

“Exogenous genetic material” is any genetic material, whether naturally occurring or otherwise, from any source that is capable of being inserted into any organism. Such exogenous genetic material includes, without limitation, nucleic acid molecules and constructs of the present invention. Exogenous genetic material may be transferred into a host cell by the use of a DNA vector or construct designed for such a purpose. Design of such a vector is generally within the skill of the art (*See, e.g., Plant Molecular Biology: A Laboratory Manual*, Clark (ed.), Springer, New York (1997)).

A construct or vector may include a promoter functional in a plant cell, or a plant promoter, to express a nucleic acid molecule of choice. A number of promoters that are active in plant cells have been described in the literature, and the CaMV 35S and FMV promoters are preferred for use in plants. Other examples of preferred promoters include bean arcelin and 7S alpha. Additional preferred promoters are enhanced or duplicated versions of the CaMV 35S and FMV 35S promoters. Odell *et al.*, *Nature* 313: 810-812 (1985); U.S. Patent No. 5,378,619. Additional promoters that may be utilized are described, for example, in U.S. Patents 5,378,619; 5,391,725; 5,428,147; 5,447,858; 5,608,144; 5,608,144; 5,614,399; 5,633,441; 5,633,435; and 4,633,436. In addition, a tissue specific enhancer may be used.

Particularly preferred promoters can also be used to express a nucleic acid molecule of the present invention in seeds or fruits. Indeed, in a preferred embodiment, the promoter used is a seed specific promoter. Examples of such promoters include the 5' regulatory regions from such genes as napin (Kridl *et al.*, *Seed Sci. Res.* 1:209-219 (1991)), phaseolin, stearyl-ACP desaturase, 7S α , 7S α ' (Chen *et al.*, *Proc. Natl. Acad. Sci.*, 83:8560-8564 (1986)), USP, arcelin and oleosin. Preferred promoters for expression in the seed are 7S α , 7S α ', napin, and *FAD2-1A* promoters.

Constructs or vectors may also include other genetic elements, including but not limited to, 3' transcriptional terminators, 3' polyadenylation signals, other untranslated nucleic acid sequences, transit or targeting sequences, selectable or screenable markers, promoters, enhancers, and operators. Constructs or vectors may also contain a promoterless gene that may utilize an endogenous promoter upon insertion.

5 Nucleic acid molecules that may be used in plant transformation or transfection may be any of the nucleic acid molecules of the present invention. It is not intended that the present invention be limited to the illustrated embodiments. Exemplary nucleic acid molecules have been described in Part A of the Detailed Description, and further non-limiting exemplary nucleic acid molecules are described below and illustrated in FIGS. 1-4, and in the Examples.

10 Referring now to the drawings, embodiments of the nucleic acid molecule of the present invention are shown in FIGS. 1-4. As described above, the nucleic acid molecule comprises (a) a first set of DNA sequences and (b) a second set of DNA sequences, which are located on one or more T-DNA regions, each of which is flanked by a right border and a left border. Within the T-DNA regions the direction of transcription is shown by arrows. The nucleic acid molecules described may have their DNA
15 sequences arranged in monocistronic or polycistronic configurations. Preferred configurations include a configuration in which the first set of DNA sequences and the second set of DNA sequences are located on a single T-DNA region.

In each of the illustrated embodiments, the first set of DNA sequences comprises one or more sequences which when expressed are capable of selectively reducing one or both of the protein and
20 transcript encoded by a gene selected from the group consisting of *FAD2*, *FAD3*, and *FATB*. Preferably each sequence in the first set of DNA sequences is capable, when expressed, of suppressing the expression of a different gene, including without limitation different gene family members. The sequences may include coding sequences, intron sequences, 3'UTR sequences, 5'UTR sequences, other non-coding sequences, or any combination of the foregoing. The first set of DNA sequences may be
25 expressed in any suitable form, including as a dsRNA construct, a sense cosuppression construct, or as an antisense construct. The first set of DNA sequences is operably linked to at least one promoter which drives expression of the sequences, which can be any promoter functional in a plant, or any plant promoter. Preferred promoters include, but are not limited to, a napin promoter, a 7S α promoter, a 7s α ' promoter, an arcelin promoter, or a *FAD2-1A* promoter.

The second set of DNA sequences comprises coding sequences, each of which is a DNA sequence that encodes a sequence that when expressed is capable of increasing one or both of the protein and transcript encoded by a gene selected from the group consisting of *KAS I*, *KAS IV*, delta-9 desaturase, and CP4 *EPSPS*. Each coding sequence is associated with a promoter, which can be any promoter functional in a plant, or any plant promoter. Preferred promoters for use in the second set of DNA sequences are an FMV promoter and/or seed-specific promoters. Particularly preferred seed-specific promoters include, but are not limited to, a napin promoter, a 7S α promoter, a 7 α ' promoter, an arcelin promoter, a delta-9 desaturase promoter, or a *FAD2-1A* promoter.

In the embodiments depicted in FIGS. 1 and 2, the first set of DNA sequences, when expressed, is capable of forming a dsRNA molecule that is capable of suppressing the expression of one or both of the protein and transcript encoded by, or transcribed from, a gene selected from the group consisting of *FAD2*, *FAD3*, and *FATB*. The first set of DNA sequences depicted in FIG. 1 comprises three non-coding sequences, each of which expresses an RNA sequence (not shown) that exhibits identity to a non-coding region of a gene selected from the group consisting of *FAD2*, *FAD3*, and *FATB* genes. The non-coding sequences each express an RNA sequence that exhibits at least 90% identity to a non-coding region of a gene selected from the group consisting of *FAD2*, *FAD3*, and *FATB* genes. The first set of DNA sequences also comprises three antisense sequences, each of which expresses an antisense RNA sequence (not shown) that is capable of forming a double-stranded RNA molecule with its respective corresponding RNA sequence (as expressed by the non-coding sequences).

The non-coding sequences may be separated from the antisense sequences by a spacer sequence, preferably one that promotes the formation of a dsRNA molecule. Examples of such spacer sequences include those set forth in Wesley *et al.*, *supra*, and Hamilton *et al.*, *Plant J.*, 15:737-746 (1988). In a preferred aspect, the spacer sequence is capable of forming a hairpin structure as illustrated in Wesley *et al.*, *supra*. Particularly preferred spacer sequences in this context are plant introns or parts thereof. A particularly preferred plant intron is a spliceable intron. Spliceable introns include, but are not limited to, an intron selected from the group consisting of PDK intron, *FAD3-1A* or *FAD3-1B* intron #5, *FAD3* intron #1, *FAD3* intron #3A, *FAD3* intron #3B, *FAD3* intron #3C, *FAD3* intron #4, *FAD3* intron #5, *FAD2* intron #1, and *FAD2-2* intron. Preferred spliceable introns include, but are not limited to, an intron selected from the group consisting of *FAD3* intron #1, *FAD3* intron #3A, *FAD3* intron #3B, *FAD3* intron

#3C, and *FAD3* intron #5. Other preferred spliceable introns include, but are not limited to, a spliceable intron that is about 0.75 kb to about 1.1 kb in length and is capable of facilitating an RNA hairpin structure. One non-limiting example of a particularly preferred spliceable intron is *FAD3* intron #5.

Referring now to FIG. 1, the nucleic acid molecule comprises two T-DNA regions, each of which is flanked by a right border and a left border. The first T-DNA region comprises the first set of DNA sequences that is operably linked to a promoter, and the first T-DNA region further comprises a first part of the second set of DNA sequences that comprises a first promoter operably linked to a first coding sequence, and a second promoter operably linked to a second coding sequence. The second T-DNA region comprises a second part of the second set of DNA sequences that comprises a third promoter operably linked to a third coding sequence. In a preferred embodiment depicted in FIG. 2, the nucleic acid molecule comprises a single T-DNA region, which is flanked by a right border and a left border. The first and second sets of DNA sequences are all located on the single T-DNA region.

In the dsRNA-expressing embodiments shown in FIGS. 1 and 2, the order of the sequences may be altered from that illustrated and described, however the non-coding sequences and the antisense sequences preferably are arranged around the spacer sequence such that, when expressed, the first non-coding sequence can hybridize to the first antisense sequence, the second non-coding sequence can hybridize to the second antisense sequence, and the third non-coding sequence can hybridize to the third antisense sequence such that a single dsRNA molecule can be formed. Preferably the non-coding sequences are in a sense orientation, and the antisense sequences are in an antisense orientation relative to the promoter. The numbers of non-coding, antisense, and coding sequences, and the various relative positions thereof on the T-DNA region(s) may also be altered in any manner suitable for achieving the goals of the present invention.

Referring now to FIGS. 3 and 4, the illustrated nucleic acid molecule comprises a T-DNA region flanked by a right border and a left border, on which are located the first and second sets of DNA sequences. The first set of DNA sequences is operably linked to a promoter and a transcriptional termination signal. The second set of DNA sequences that comprises a first promoter operably linked to a first coding sequence, a second promoter operably linked to a second coding sequence, and a third promoter operably linked to a third coding sequence. The transcriptional termination signal can be any transcriptional termination signal functional in a plant, or any plant transcriptional termination signal.

Preferred transcriptional termination signals include, but are not limited to, a pea Rubisco E9 3' sequence, a *Brassica napin* 3' sequence, a *tml* 3' sequence, and a *nos* 3' sequence.

In the embodiment depicted in FIG. 3, the first set of DNA sequences, when expressed, is capable of forming a sense cosuppression construct that is capable of suppressing the expression of one or more proteins or transcripts encoded by, or derived from, a gene selected from the group consisting of *FAD2*, *FAD3*, and *FATB*. The first set of DNA sequences comprises three non-coding sequences, each of which expresses an RNA sequence (not shown) that exhibits identity to one or more non-coding region(s) of a gene selected from the group consisting of *FAD2*, *FAD3*, and *FATB* genes. The non-coding sequences each express an RNA sequence that exhibits at least 90% identity to one or more non-coding region(s) of a gene selected from the group consisting of *FAD2*, *FAD3*, and *FATB* genes. The order of the non-coding sequences within the first set of DNA sequences may be altered from that illustrated and described herein, but the non-coding sequences are arranged in a sense orientation relative to the promoter.

FIG. 4 depicts an embodiment in which the first set of DNA sequences, when expressed, is capable of forming an antisense construct that is capable of suppressing the expression of one or more proteins or transcripts encoded by, or derived from, a gene selected from the group consisting of *FAD2*, *FAD3*, and *FATB*. The first set of DNA sequences comprises three antisense sequences, each of which expresses an antisense RNA sequence (not shown) that exhibits identity to one or more non-coding region(s) of a gene selected from the group consisting of *FAD2*, *FAD3*, and *FATB* genes. The antisense sequences each express an antisense RNA sequence that exhibits at least 90% identity to one or more non-coding region(s) of a gene selected from the group consisting of *FAD2*, *FAD3*, and *FATB* genes. The order of the antisense sequences within the first set of DNA sequences may be altered from that illustrated and described herein, but the antisense sequences are arranged in an antisense orientation relative to the promoter.

The above-described nucleic acid molecules are preferred embodiments which achieve the objects, features and advantages of the present invention. It is not intended that the present invention be limited to the illustrated embodiments. The arrangement of the sequences in the first and second sets of DNA sequences within the nucleic acid molecule is not limited to the illustrated and described arrangements, and may be altered in any manner suitable for achieving the objects, features and advantages of the present invention as described herein and illustrated in the accompanying drawings.

E. Transgenic Organisms, and Methods for Producing Same

Any of the nucleic acid molecules and constructs of the invention may be introduced into a plant or plant cell in a permanent or transient manner. Preferred nucleic acid molecules and constructs of the present invention are described above in Parts A through D of the Detailed Description, and in the
5 Examples. Another embodiment of the invention is directed to a method of producing transgenic plants which generally comprises the steps of selecting a suitable plant or plant cell, transforming the plant or plant cell with a recombinant vector, and obtaining a transformed host cell.

In a preferred embodiment the plant or cell is, or is derived from, a plant involved in the production of vegetable oils for edible and industrial uses. Especially preferred are temperate oilseed
10 crops. Plants of interest include, but are not limited to, rapeseed (canola and High Erucic Acid varieties), maize, soybean, crambe, mustard, castor bean, peanut, sesame, cotton, linseed, safflower, oil palm, flax, sunflower, and coconut. The invention is applicable to monocotyledonous or dicotyledonous species alike, and will be readily applicable to new and/or improved transformation and regulatory techniques.

Methods and technology for introduction of DNA into plant cells are well known to those of skill
15 in the art, and virtually any method by which nucleic acid molecules may be introduced into a cell is suitable for use in the present invention. Non-limiting examples of suitable methods include: chemical methods; physical methods such as microinjection, electroporation, the gene gun, microprojectile bombardment, and vacuum infiltration; viral vectors; and receptor-mediated mechanisms. Other methods of cell transformation can also be used and include but are not limited to introduction of DNA into plants
20 by direct DNA transfer into pollen, by direct injection of DNA into reproductive organs of a plant, or by direct injection of DNA into the cells of immature embryos followed by the rehydration of desiccated embryos.

Agrobacterium-mediated transfer is a widely applicable system for introducing genes into plant cells. See, e.g., Fraley *et al.*, *Bio/Technology* 3:629-635 (1985); Rogers *et al.*, *Methods Enzymol.*
25 153:253-277 (1987). The region of DNA to be transferred is defined by the border sequences and intervening DNA is usually inserted into the plant genome. Spielmann *et al.*, *Mol. Gen. Genet.* 205:34 (1986). Modern *Agrobacterium* transformation vectors are capable of replication in *E. coli* as well as *Agrobacterium*, allowing for convenient manipulations. Klee *et al.*, In: *Plant DNA Infectious Agents*, Hohn and Schell (eds.), Springer-Verlag, New York, pp. 179-203 (1985).

The regeneration, development and cultivation of plants from single plant protoplast transformants or from various transformed explants is well known in the art. *See generally*, Maliga *et al.*, *Methods in Plant Molecular Biology*, Cold Spring Harbor Press (1995); Weissbach and Weissbach, In: *Methods for Plant Molecular Biology*, Academic Press, San Diego, CA (1988). Plants of the present invention can be part of or generated from a breeding program, and may also be reproduced using apomixis. Methods for the production of apomictic plants are known in the art. *See, e.g.*, U.S. Patent 5,811,636.

In a preferred embodiment, a plant of the present invention that includes nucleic acid sequences which when expressed are capable of selectively reducing the level of a FAD2, FAD3, and/or FATB protein, and/or a FAD2, FAD3, and/or FATB transcript is mated with another plant of the present invention that includes nucleic acid sequences which when expressed are capable of overexpressing another enzyme. Preferably the other enzyme is selected from the group consisting of beta-ketoacyl-ACP synthase I, beta-ketoacyl-ACP synthase IV, delta-9 desaturase, and CP4 EPSPS.

F. Products of the Present Invention

The plants of the present invention may be used in whole or in part. Preferred plant parts include reproductive or storage parts. The term "plant parts" as used herein includes, without limitation, seed, endosperm, ovule, pollen, roots, tubers, stems, leaves, stalks, fruit, berries, nuts, bark, pods, seeds and flowers. In a particularly preferred embodiment of the present invention, the plant part is a seed.

Any of the plants or parts thereof of the present invention may be processed to produce a feed, meal, protein, or oil preparation. A particularly preferred plant part for this purpose is a seed. In a preferred embodiment the feed, meal, protein or oil preparation is designed for livestock animals, fish or humans, or any combination. Methods to produce feed, meal, protein and oil preparations are known in the art. *See, e.g.*, U.S. Patents 4,957,748, 5,100,679, 5,219,596, 5,936,069, 6,005,076, 6,146,669, and 6,156,227. In a preferred embodiment, the protein preparation is a high protein preparation. Such a high protein preparation preferably has a protein content of greater than 5% w/v, more preferably 10% w/v, and even more preferably 15% w/v.

In a preferred oil preparation, the oil preparation is a high oil preparation with an oil content derived from a plant or part thereof of the present invention of greater than 5% w/v, more preferably 10% w/v, and even more preferably 15% w/v. In a preferred embodiment the oil preparation is a liquid and of

a volume greater than 1, 5, 10 or 50 liters. The present invention provides for oil produced from plants of the present invention or generated by a method of the present invention. Such an oil may exhibit enhanced oxidative stability. Also, such oil may be a minor or major component of any resultant product.

Moreover, such oil may be blended with other oils. In a preferred embodiment, the oil produced from plants of the present invention or generated by a method of the present invention constitutes greater than 0.5%, 1%, 5%, 10%, 25%, 50%, 75% or 90% by volume or weight of the oil component of any product. In another embodiment, the oil preparation may be blended and can constitute greater than 10%, 25%, 35%, 50% or 75% of the blend by volume. Oil produced from a plant of the present invention can be admixed with one or more organic solvents or petroleum distillates.

Seeds of the plants may be placed in a container. As used herein, a container is any object capable of holding such seeds. A container preferably contains greater than about 500, 1,000, 5,000, or 25,000 seeds where at least about 10%, 25%, 50%, 75% or 100% of the seeds are derived from a plant of the present invention. The present invention also provides a container of over about 10,000, more preferably about 20,000, and even more preferably about 40,000 seeds where over about 10%, more preferably about 25%, more preferably 50% and even more preferably about 75% or 90% of the seeds are seeds derived from a plant of the present invention. The present invention also provides a container of over about 10 kg, more preferably about 25 kg, and even more preferably about 50 kg seeds where over about 10%, more preferably about 25%, more preferably about 50% and even more preferably about 75% or 90% of the seeds are seeds derived from a plant of the present invention.

G. Oil Compositions

For many oil applications, saturated fatty acid levels are preferably less than 8% by weight, and more preferably about 2-3% by weight. Saturated fatty acids have high melting points which are undesirable in many applications. When used as a feedstock for fuel, saturated fatty acids cause clouding at low temperatures, and confer poor cold flow properties such as pour points and cold filter plugging points to the fuel. Oil products containing low saturated fatty acid levels may be preferred by consumers and the food industry because they are perceived as healthier and/or may be labeled as "saturated fat free" in accordance with FDA guidelines. In addition, low saturate oils reduce or eliminate the need to winterize the oil for food applications such as salad oils. In biodiesel and lubricant applications oils with low saturated fatty acid levels confer improved cold flow properties and do not cloud at low temperatures.

The factors governing the physical properties of a particular oil are complex. Palmitic, stearic and other saturated fatty acids are typically solid at room temperature, in contrast to the unsaturated fatty acids, which remain liquid. Because saturated fatty acids have no double bonds in the acyl chain, they remain stable to oxidation at elevated temperatures. Saturated fatty acids are important components in margarines and chocolate formulations, but for many food applications, reduced levels of saturated fatty acids are desired.

Oleic acid has one double bond, but is still relatively stable at high temperatures, and oils with high levels of oleic acid are suitable for cooking and other processes where heating is required. Recently, increased consumption of high oleic oils has been recommended, because oleic acid appears to lower blood levels of low density lipoproteins ("LDLs") without affecting levels of high density lipoproteins ("HDLs"). However, some limitation of oleic acid levels is desirable, because when oleic acid is degraded at high temperatures, it creates negative flavor compounds and diminishes the positive flavors created by the oxidation of linoleic acid. Neff *et al.*, *JAOCS*, 77 :1303-1313 (2000); Warner *et al.*, *J. Agric. Food Chem.* 49:899-905 (2001). Preferred oils have oleic acid levels that are 65-85% or less by weight, in order to limit off-flavors in food applications such as frying oil and fried food. Other preferred oils have oleic acid levels that are greater than 55% by weight in order to improve oxidative stability.

Linoleic acid is a major polyunsaturated fatty acid in foods and is an essential nutrient for humans. It is a desirable component for many food applications because it is a major precursor of fried food flavor substances such as 2,4 decadienal, which make fried foods taste good. However, linoleic acid has limited stability when heated. Preferred food oils have linoleic acid levels that are 10% or greater by weight, to enhance the formation of desirable fried food flavor substances, and also are 25% or less by weight, so that the formation of off-flavors is reduced. Linoleic acid also has cholesterol-lowering properties, although dietary excess can reduce the ability of human cells to protect themselves from oxidative damage, thereby increasing the risk of cardiovascular disease. Toborek *et al.*, *Am J. Clin. J.* 75:119-125 (2002). See generally *Flavor Chemistry of Lipid Foods*, editors D.B. Min & T.H. Smouse, Am Oil Chem. Soc., Champaign, IL (1989).

Linoleic acid, having a lower melting point than oleic acid, further contributes to improved cold flow properties desirable in biodiesel and biolubricant applications. Preferred oils for most applications have linoleic acid levels of 30% or less by weight, because the oxidation of linoleic acid limits the useful

storage or use-time of frying oil, food, feed, fuel and lubricant products. *See generally, Physical Properties of Fats, Oils, and Emulsifiers*, ed. N. Widlak, AOCS Press (1999); Erhan & Asadauskas, *Lubricant Basestocks from Vegetable Oils, Industrial Crops and Products*, 11:277-282 (2000). In addition, high linoleic acid levels in cattle feed can lead to undesirably high levels of linoleic acid in the milk of dairy cattle, and therefore poor oxidative stability and flavor. Timmons *et al.*, *J. Dairy Sci.* 84:2440-2449 (2001). A broadly useful oil composition has linoleic acid levels of 10-25% by weight.

Linolenic acid is also an important component of the human diet. It is used to synthesize the ω -3 family of long-chain fatty acids and the prostaglandins derived therefrom. However, its double bonds are highly susceptible to oxidation, so that oils with high levels of linolenic acid deteriorate rapidly on exposure to air, especially at high temperatures. Partial hydrogenation of such oils is often necessary before they can be used in food products to retard the formation of off-flavors and rancidity when the oil is heated, but hydrogenation creates unhealthy *trans* fatty acids which can contribute to cardiovascular disease. To achieve improved oxidative stability, and reduce the need to hydrogenate oil, preferred oils have linolenic acid levels that are 8% or less by weight, 6% or less, 4% or less, and more preferably 0.5-2% by weight of the total fatty acids in the oil of the present invention.

The oil of the present invention can be a blended oil, synthesized oil or in a preferred embodiment an oil generated from an oilseed having an appropriate oil composition. In a preferred embodiment, the oil is a soybean oil. The oil can be a crude oil such as crude soybean oil, or can be a processed oil, for example the oil can be refined, bleached, deodorized, and/or winterized. As used herein, "refining" refers to a process of treating natural or processed fat or oil to remove impurities, and may be accomplished by treating fat or oil with caustic soda, followed by centrifugation, washing with water, and heating under vacuum. "Bleaching" refers to a process of treating a fat or oil to remove or reduce the levels of coloring materials in the fat or oil. Bleaching may be accomplished by treating fat or oil with activated charcoal or Fullers (diatomaceous) earth. "Deodorizing" refers to a process of removing components from a fat or oil that contribute objectionable flavors or odors to the end product, and may be accomplished by use of high vacuum and superheated steam washing. "Winterizing" refers to a process of removing saturated glycerides from an oil, and may be accomplished by chilling and removal of solidified portions of fat from an oil.

A preferred oil of the present invention has a low saturate oil composition, or a zero saturate oil composition. In other preferred embodiments, oils of the present invention have increased oleic acid levels, reduced saturated fatty levels, and reduced polyunsaturated fatty acid levels. In a preferred embodiment, the oil is a soybean oil. The percentages of fatty acid content, or fatty acid levels, used
5 herein refer to percentages by weight.

In a first embodiment, an oil of the present invention preferably has an oil composition that is 55 to 80% oleic acid, 10 to 40% linoleic acid, 6% or less linolenic acid, and 2 to 8% saturated fatty acids; more preferably has an oil composition that is 55 to 80% oleic acid, 10 to 39% linoleic acid, 4.5% or less linolenic acid, and 3 to 6% saturated fatty acids; and even more preferably has an oil composition that is
10 55 to 80% oleic acid, 10 to 39% linoleic acid, 3.0% or less linolenic acid, and 2 to 3.6% saturated fatty acids.

In a second embodiment, an oil of the present invention preferably has an oil composition that is 65 to 80% oleic acid, 10 to 30% linoleic acid, 6% or less linolenic acid, and 2 to 8% saturated fatty acids; more preferably has an oil composition that is 65 to 80% oleic acid, 10 to 29% linoleic acid, 4.5% or less
15 linolenic acid, and 3 to 6% saturated fatty acids; and even more preferably has an oil composition that is 65 to 80% oleic acid, 10 to 29% linoleic acid, 3.0% or less linolenic acid, and 2 to 3.6% saturated fatty acids.

In another embodiment, an oil of the present invention has an oil composition that is about 65-80% oleic acid, about 3-8% saturates, and about 10-20% polyunsaturates. In another embodiment, an oil
20 of the present invention has an oil composition that is about 65-80% oleic acid, about 2-3.5% saturates; and about 10-25% polyunsaturates.

In other embodiments, the percentage of oleic acid is 50% or greater; 55% or greater; 60% or greater; 65% or greater; 70% or greater; 75% or greater; or 80% or greater; or is a range from 50 to 80%; 55 to 80%; 55 to 75%; 55 to 65%; 65 to 80%; 65 to 75%; 65 to 70%; or 69 to 73%. Suitable percentage
25 ranges for oleic acid content in oils of the present invention also include ranges in which the lower limit is selected from the following percentages: 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, or 80 percent; and the upper limit is selected from the following percentages: 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, or 90 percent.

In these other embodiments, the percentage of linoleic acid in an oil of the present invention is a range from 10 to 40%; 10 to 39%; 10 to 30%; 10 to 29%; 10 to 28%; 10 to 25%; 10 to 21%; 10 to 20%; 11 to 30%; 12 to 30%; 15 to 25%; 20 to 25%; 20 to 30%; or 21 to 24%. Suitable percentage ranges for linoleic acid content in oils of the present invention also include ranges in which the lower limit is
5 selected from the following percentages: 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, or 30 percent; and the upper limit is selected from the following percentages: 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, or 40 percent.

In these other embodiments, the percentage of linolenic acid in an oil of the present invention is 10% or less; 9% or less; 8% or less; 7% or less; 6% or less; 5% or less; 4.5% or less; 4% or less; 3.5% or
10 less; 3% or less; 3.0% or less; 2.5% or less; or 2% or less; or is a range from 0.5 to 2%; 0.5 to 3%; 0.5 to 4.5%; 0.5% to 6%; 3 to 5%; 3 to 6%; 3 to 8%; 1 to 2%; 1 to 3%; or 1 to 4%. In these other embodiments, the percentage of saturated fatty acids in an oil composition of the present invention is 15% or less; 14% or less; 13% or less; 12% or less, 11% or less; 10% or less; 9% or less; 8% or less; 7% or less; 6% or less; 5% or less; 4% or less; or 3.6% or less; or is a range from 2 to 3%; 2 to 3.6%; 2 to 4%; 2 to 8%; 3 to 15%;
15 3 to 10%; 3 to 8%; 3 to 6%; 3.6 to 7%; 5 to 8%; 7 to 10%; or 10 to 15%.

An oil of the present invention is particularly suited to use as a cooking or frying oil. Because of its reduced polyunsaturated fatty acid content, the oil of the present invention does not require the extensive processing of typical oils because fewer objectionable odorous and colorant compounds are present. In addition, the low saturated fatty acid content of the present oil improves the cold flow
20 properties of the oil, and obviates the need to heat stored oil to prevent it from crystallizing or solidifying. Improved cold flow also enhances drainage of oil from fried food material once it has been removed from frying oil, thereby resulting in a lower fat product. See Bouchon *et al.*, *J. Food Science* 66: 918-923 (2001). The low levels of linolenic acid in the present oil are particularly advantageous in frying to reduce off-flavors.

The present oil is also well-suited for use as a salad oil (an oil that maintains clarity at refrigerator temperatures of 40-50 degrees Fahrenheit). Its improved clarity at refrigerator temperatures, due to its low saturated fatty acid and moderate linoleic acid content, reduces or eliminates the need to winterize the oil before use as a salad oil.

In addition, the moderate linoleic and low linolenic acid content of the present oil make it well-suited for the production of shortening, margarine and other semi-solid vegetable fats used in foodstuffs. Production of these fats typically involves hydrogenation of unsaturated oils such as soybean oil, corn oil, or canola oil. The increased oxidative and flavor stability of the present oil mean that it need not be
5 hydrogenated to the extent that typical vegetable oil is for uses such as margarine and shortening, thereby reducing processing costs and the production of unhealthy *trans* isomers.

An oil of the present invention is also suitable for use as a feedstock to produce biodiesel, particularly because biodiesel made from such an oil has improved cold flow, improved ignition quality (cetane number), improved oxidative stability, and reduced nitric oxide emissions. Biodiesel is an
10 alternative diesel fuel typically comprised of methyl esters of saturated, monounsaturated, and polyunsaturated C₁₆-C₂₂ fatty acids. Cetane number is a measure of ignition quality – the shorter the ignition delay time of fuel in the engine, the higher the cetane number. The ASTM standard specification for biodiesel fuel (D 6751-02) requires a minimum cetane number of 47.

The use of biodiesel in conventional diesel engines results in substantial reductions of pollutants
15 such as sulfates, carbon monoxide, and particulates compared to petroleum diesel fuel, and use in school buses can greatly reduce children's exposure to toxic diesel exhaust. A limitation to the use of 100% conventional biodiesel as fuel is the high cloud point of conventional soy biodiesel (2 degrees C) compared to number 2 diesel (-16 degrees C). Dunn *et al.*, *Recent. Res. Devel. in Oil Chem.*, 1:31-56 (1997). Biodiesel made from oil of the present invention has an improved (reduced) cloud point and cold
20 filter plugging point, and may also be used in blends to improve the cold-temperature properties of biodiesel made from inexpensive but highly saturated sources of fat such as animal fats (tallow, lard, chicken fat) and palm oil. Biodiesel can also be blended with petroleum diesel at any level.

Biodiesel is typically obtained by extracting, filtering and refining soybean oil to remove free fats and phospholipids, and then transesterifying the oil with methanol to form methyl esters of the fatty acids.
25 *See, e.g.*, U.S. Patent No. 5,891,203. The resultant soy methyl esters are commonly referred to as "biodiesel." The oil of the present invention may also be used as a diesel fuel without the formation of methyl esters, such as, for example, by mixing acetals with the oil. *See, e.g.*, U.S. Patent No. 6,013,114. Due to its improved cold flow and oxidative stability properties, the oil of the present invention is also

useful as a lubricant, and as a diesel fuel additive. *See, e.g.*, U.S. Patent Nos. 5,888,947, 5,454,842 and 4,557,734.

Soybeans and oils of the present invention are also suitable for use in a variety of soyfoods made from whole soybeans, such as soymilk, soy nut butter, natto, and tempeh, and soyfoods made from processed soybeans and soybean oil, including soybean meal, soy flour, soy protein concentrate, soy protein isolates, texturized soy protein concentrate, hydrolyzed soy protein, whipped topping, cooking oil, salad oil, shortening, and lecithin. Whole soybeans are also edible, and are typically sold to consumers raw, roasted, or as edamamé. Soymilk, which is typically produced by soaking and grinding whole soybeans, may be consumed as is, spray-dried, or processed to form soy yogurt, soy cheese, tofu, or yuba.

5

10 The present soybean or oil may be advantageously used in these and other soyfoods because of its improved oxidative stability, the reduction of off-flavor precursors, and its low saturated fatty acid level.

The following examples are illustrative and not intended to be limiting in any way.

All publications, patents, and patent applications mentioned in this specification are herein incorporated by reference to the same extent as if each individual publication, patent, or patent application was specifically and individually indicated to be incorporated by reference.

15

EXAMPLES

Example 1 Isolation of *FATB-2* Sequences

Leaf tissue is obtained from Asgrow soy variety A3244, ground in liquid nitrogen and stored at -80°C until use. Six ml of SDS Extraction buffer (650 ml sterile ddH₂O, 100 ml 1M Tris-Cl pH 8, 100 ml 0.25M EDTA, 50 ml 20% SDS, 100 ml 5M NaCl, 4 µl beta-mercaptoethanol) is added to 2 ml of frozen/ground leaf tissue, and the mixture is incubated at 65°C for 45 minutes. The sample is shaken every 15 minutes. 2 ml of ice-cold 5M potassium acetate is added to the sample, the sample is shaken, and then is incubated on ice for 20 minutes. 3 ml of CHCl₃ is added to the sample and the sample is shaken for 10 minutes.

20

The sample is centrifuged at 10,000 rpm for 20 minutes and the supernatant is collected. 2 ml of isopropanol is added to the supernatant and mixed. The sample is then centrifuged at 10,000 rpm for 20 minutes and the supernatant is drained. The pellet is resuspended in 200 µl RNase and incubated at 65°C

25

for 20 minutes. 300 µl ammonium acetate/isopropanol (1:7) is added and mixed. The sample is then centrifuged at 10,000 rpm for 15 minutes and the supernatant is discarded. The pellet is rinsed with 500 µl 80% ethanol and allowed to air dry. The pellet of genomic DNA is then resuspended in 200 µl T10E1 (10mM Tris:1mM EDTA).

5 A soy *FATB-2* cDNA contig sequence (SEQ ID NO: 42) is used to design thirteen oligonucleotides that span the gene: F1 (SEQ ID NO: 48), F2 (SEQ ID NO: 49), F3 (SEQ ID NO: 50), F4 (SEQ ID NO: 51), F5 (SEQ ID NO: 52), F6 (SEQ ID NO: 53), F7 (SEQ ID NO: 54), R1 (SEQ ID NO: 55), R2 (SEQ ID NO: 56), R3 (SEQ ID NO: 57), R4 (SEQ ID NO: 58), R5 (SEQ ID NO: 59), and R6 (SEQ ID NO: 60). The oligonucleotides are used in pairs for PCR
10 amplification from the isolated soy genomic DNA: pair 1 (F1 + R1), pair 2 (F2 + R1), pair 3 (F3 + R2), pair 4 (F4 + R3), pair 5 (F5 + R4), pair 6 (F6 + R5), and pair 7 (F7 + R6). The PCR amplification for pair 5 is carried out as follows: 1 cycle, 95°C for 10 minutes; 30 cycles, 95°C for 15 sec, 43°C for 30 sec, 72°C for 45 sec; 1 cycle, 72°C for 7 minutes. For all other oligo
15 pairs, PCR amplifications are carried out as follows: 1 cycle, 95°C for 10 minutes; 30 cycles, 95°C for 15 sec, 48°C for 30 sec, 72°C for 45 sec; 1 cycle, 72°C for 7 minutes. Positive fragments are obtained from primer pairs 1, 2, 4, 5, 6 and 7. Each fragment is cloned into vector pCR2.1 (Invitrogen). Fragments 2, 4, 5 and 6 are confirmed and sequenced. These four sequences are aligned to form a genomic sequence for the *FATB-2* gene (SEQ ID NO: 43).

Four introns are identified in the soybean *FATB-2* gene by comparison of the genomic sequence
20 to the cDNA sequence: intron I (SEQ ID NO: 44) spans base 119 to base 1333 of the genomic sequence (SEQ ID NO: 43) and is 1215 bp in length; intron II (SEQ ID NO: 45) spans base 2231 to base 2568 of the genomic sequence (SEQ ID NO: 43) and is 338 bp in length; intron III (SEQ ID NO: 46) spans base 2702 to base 3342 of the genomic sequence (SEQ ID NO: 43) and is 641 bp in length; and intron IV (SEQ ID NO: 47) spans base 3457 to base 3823 of the genomic sequence (SEQ ID NO: 43) and is 367 bp
25 in length.

Example 2 Suppression Constructs

2A. *FAD2-1* Constructs

The *FAD2-1A* intron #1 (SEQ ID NO: 1) is cloned into the expression cassette, pCGN3892, in sense and antisense orientations. The vector pCGN3892 contains the soybean 7S promoter and a pea *rbcS* 3'. Both gene fusions are then separately ligated into pCGN9372, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter. The resulting expression constructs (pCGN5469 sense and pCGN5471 antisense) are used for transformation of soybean.

The *FAD2-1B* intron (SEQ ID NO: 2) is fused to the 3' end of the *FAD2-1A* intron #1 in plasmid pCGN5468 (contains the soybean 7S promoter fused to the *FAD2-1A* intron (sense) and a pea *rbcS* 3') or pCGN5470 (contains the soybean 7S promoter fused to the *FAD2-1A* intron (antisense) and a pea *rbcS* 3') in sense or antisense orientation, respectively. The resulting intron combination fusions are then ligated separately into pCGN9372, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter. The resulting expression constructs (pCGN5485, *FAD2-1A* & *FAD2-1B* intron sense and pCGN5486, *FAD2-1A* & *FAD2-1B* intron antisense) are used for transformation of soybean.

2B. *FAD3-1* Constructs

FAD3-1A introns #1, #2, #4 and #5 (SEQ ID NOs: 7, 8, 10 and 11, respectively), *FAD3-1B* introns #3C (SEQ ID NO: 23) and #4 (SEQ ID NO: 24), are all ligated separately into pCGN3892, in sense or antisense orientations. pCGN3892 contains the soybean 7S promoter and a pea *rbcS* 3'. These fusions are ligated into pCGN9372, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter for transformation into soybean. The resulting expression constructs (pCGN5455, *FAD3-1A* intron #4 sense; pCGN5459, *FAD3-1A* intron #4 antisense; pCGN5456, *FAD3* intron #5 sense; pCGN5460, *FAD3-1A* intron #5 antisense; pCGN5466, *FAD3-1A* intron #2 antisense; pCGN5473, *FAD3-1A* intron #1 antisense) are used for transformation of soybean.

2C. *FatB* Constructs

The soybean *FATB-1* intron II sequence (SEQ ID NO: 30) is amplified via PCR using a *FATB-1* partial genomic clone as a template. PCR amplification is carried out as follows: 1 cycle, 95°C for 10 min; 25 cycles, 95°C for 30 sec, 62°C for 30 sec, 72°C for 30 sec; 1 cycle, 72°C for 7 min. PCR

amplification results in a product that is 854 bp long, including reengineered restriction sites at both ends. The PCR product is cloned directly into the expression cassette pCGN3892 in sense orientation, by way of *XhoI* sites engineered onto the 5' ends of the PCR primers, to form pMON70674. Vector pCGN3892 contains the soybean 7S promoter and a pea *rbcS* 3'. pMON70674 is then cut with *NotI* and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter. The resulting gene expression construct, pMON70678, is used for transformation of soybean using *Agrobacterium* methods.

Two other expression constructs containing the soybean *FATB-I* intron II sequence (SEQ ID NO: 30) are created. pMON70674 is cut with *NotI* and ligated into pMON70675 which contains the CP4 *EPSPS* gene regulated by the FMV promoter and the *KAS IV* gene regulated by the napin promoter, resulting in pMON70680. The expression vector pMON70680 is then cut with *SnaBI* and ligated with a gene fusion of the jojoba delta-9 desaturase gene (SEQ ID NO: 41) in sense orientation regulated by the 7S promoter. The expression constructs pMON70680 and pMON70681 are used for transformation of soybean using *Agrobacterium* methods.

15 2D Combination Constructs

Expression constructs are made containing various permutations of a first set of DNA sequences. The first set of DNA sequences are any of those described, or illustrated in FIGS. 5 and 6, or any other set of DNA sequences that contain either various combinations of sense and antisense *FAD2*, *FAD3*, and *FATB* non-coding regions so that they are capable of forming dsRNA constructs, sense cosuppression constructs, antisense constructs, or various combinations of the foregoing.

FIGS. 5(a)-(c) depict several first sets of DNA sequences which are capable of expressing sense cosuppression or antisense constructs according to the present invention, the non-coding sequences of which are described in Tables 1 and 2 below. The non-coding sequences may be single sequences, combinations of sequences (*e.g.*, the 5'UTR linked to the 3'UTR), or any combination of the foregoing. To express a sense cosuppression construct, all of the non-coding sequences are sense sequences, and to express an antisense construct, all of the non-coding sequences are antisense sequences. FIG. 5(d) depicts a first set of DNA sequences which is capable of expressing sense and antisense constructs according to the present invention.

FIGS. 6(a)-(c) depict several first sets of DNA sequences which are capable of expressing dsRNA constructs according to the present invention, the non-coding sequences of which are described in Tables 1 and 2 below. The first set of DNA sequences depicted in FIG. 6 comprises pairs of related sense and antisense sequences, arranged such that, *e.g.*, the RNA expressed by the first sense sequence is capable of forming a double-stranded RNA with the antisense RNA expressed by the first antisense sequence. For example, referring to FIG. 6(a) and illustrative combination No. 1 (of Table 1), the first set of DNA sequences comprises a sense *FAD2-1* sequence, a sense *FAD3-1* sequence, an antisense *FAD2-1* sequence and an antisense *FAD3-1* sequence. Both antisense sequences correspond to the sense sequences so that the expression products of the first set of DNA sequences are capable of forming a double-stranded RNA with each other. The sense sequences may be separated from the antisense sequences by a spacer sequence, preferably one that promotes the formation of a dsRNA molecule. Examples of such spacer sequences include those set forth in Wesley *et al.*, *supra*, and Hamilton *et al.*, *Plant J.*, 15:737-746 (1988). The promoter is any promoter functional in a plant, or any plant promoter. Non-limiting examples of suitable promoters are described in Part D of the Detailed Description.

The first set of DNA sequences is inserted in an expression construct in either the sense or antisense orientation using a variety of DNA manipulation techniques. If convenient restriction sites are present in the DNA sequences, they are inserted into the expression construct by digesting with the restriction endonucleases and ligation into the construct that has been digested at one or more of the available cloning sites. If convenient restriction sites are not available in the DNA sequences, the DNA of either the construct or the DNA sequences is modified in a variety of ways to facilitate cloning of the DNA sequences into the construct. Examples of methods to modify the DNA include by PCR, synthetic linker or adapter ligation, in vitro site-directed mutagenesis, filling in or cutting back of overhanging 5' or 3' ends, and the like. These and other methods of manipulating DNA are well known to those of ordinary skill in the art.

Table 1

Illustrative Combinations	Non-Coding Sequences (sense or antisense)			
	First	Second	Third	Fourth
1	<i>FAD2-1A</i> or <i>B</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>		
2	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FAD2-1A</i> or <i>B</i>		
3	<i>FAD2-1A</i> or <i>B</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence	
4	<i>FAD2-1A</i> or <i>B</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FATB-1</i>	
5	<i>FAD2-1A</i> or <i>B</i>	<i>FATB-1</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	
6	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FAD2-1A</i> or <i>B</i>	<i>FATB-1</i>	
7	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FATB-1</i>	<i>FAD2-1A</i> or <i>B</i>	
8	<i>FATB-1</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FAD2-1A</i> or <i>B</i>	
9	<i>FATB-1</i>	<i>FAD2-1A</i> or <i>B</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	
10	<i>FAD2-1A</i> or <i>B</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence	<i>FATB-1</i>
11	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FAD2-1A</i> or <i>B</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence	<i>FATB-1</i>
12	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence	<i>FAD2-1A</i> or <i>B</i>	<i>FATB-1</i>
13	<i>FAD2-1A</i> or <i>B</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FATB-1</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence
14	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FAD2-1A</i> or <i>B</i>	<i>FATB-1</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence
15	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence	<i>FATB-1</i>	<i>FAD2-1A</i> or <i>B</i>
16	<i>FAD2-1A</i> or <i>B</i>	<i>FATB-1</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence
17	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FATB-1</i>	<i>FAD2-1A</i> or <i>B</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence
18	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FATB-1</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence	<i>FAD2-1A</i> or <i>B</i>
19	<i>FATB-1</i>	<i>FAD2-1A</i> or <i>B</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence
20	<i>FATB-1</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FAD2-1A</i> or <i>B</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence
21	<i>FATB-1</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence	<i>FAD2-1A</i> or <i>B</i>
22	<i>FAD2-1A</i> or <i>B</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FATB-2</i>	
23	<i>FAD2-1A</i> or <i>B</i>	<i>FATB-2</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	
24	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FAD2-1A</i> or <i>B</i>	<i>FATB-2</i>	
25	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FATB-2</i>	<i>FAD2-1A</i> or <i>B</i>	
26	<i>FATB-2</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FAD2-1A</i> or <i>B</i>	
27	<i>FATB-2</i>	<i>FAD2-1A</i> or <i>B</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	
28	<i>FAD2-1A</i> or <i>B</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence	<i>FATB-2</i>
29	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FAD2-1A</i> or <i>B</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence	<i>FATB-2</i>
30	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence	<i>FAD2-1A</i> or <i>B</i>	<i>FATB-2</i>
31	<i>FAD2-1A</i> or <i>B</i>	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FATB-2</i>	different <i>FAD3-1A</i> or <i>B</i> or <i>C</i> sequence
32	<i>FAD3-1A</i> or <i>B</i> or <i>C</i>	<i>FAD2-1A</i> or <i>B</i>	<i>FATB-2</i>	different <i>FAD3-1A</i> or <i>B</i>

Illustrative Combinations	Non-Coding Sequences (sense or antisense)			
	First	Second	Third	Fourth
				or C sequence
33	<i>FAD3-1A</i> or B or C	different <i>FAD3-1A</i> or B or C sequence	<i>FATB-2</i>	<i>FAD2-1A</i> or B
34	<i>FAD2-1A</i> or B	<i>FATB-2</i>	<i>FAD3-1A</i> or B or C	different <i>FAD3-1A</i> or B or C sequence
35	<i>FAD3-1A</i> or B or C	<i>FATB-2</i>	<i>FAD2-1A</i> or B	different <i>FAD3-1A</i> or B or C sequence
36	<i>FAD3-1A</i> or B or C	<i>FATB-2</i>	different <i>FAD3-1A</i> or B or C sequence	<i>FAD2-1A</i> or B
37	<i>FATB-2</i>	<i>FAD2-1A</i> or B	<i>FAD3-1A</i> or B or C	different <i>FAD3-1A</i> or B or C sequence
38	<i>FATB-2</i>	<i>FAD3-1A</i> or B or C	<i>FAD2-1A</i> or B	different <i>FAD3-1A</i> or B or C sequence
39	<i>FATB-2</i>	<i>FAD3-1A</i> or B or C	different <i>FAD3-1A</i> or B or C sequence	<i>FAD2-1A</i> or B

Table 2

	Correlation of SEQ ID NOs with Sequences in Table 1						
	<i>FAD2-1A</i>	<i>FAD2-1B</i>	<i>FAD3-1A</i>	<i>FAD3-1B</i>	<i>FAD3-1C</i>	<i>FATB-1</i>	<i>FATB-2</i>
3'UTR	SEQ NO: 5	n/a	SEQ NO: 16	SEQ NO: 26	n/a	SEQ NO: 36	n/a
5'UTR	SEQ NO: 6	n/a	SEQ NO: 17	SEQ NO: 27	n/a	SEQ NO: 37	n/a
5'+3' UTR (or 3'+5' UTR)	Linked SEQ NOs: 5 and 6	n/a	Linked SEQ NOs: 16 and 17	Linked SEQ NOs: 26 and 27	n/a	Linked SEQ NOs: 36 and 37	n/a
Intron #1	SEQ NO: 1	SEQ NO: 2	SEQ NO: 7	SEQ NO: 19	n/a	SEQ NO: 29	SEQ NO: 44
Intron #2	n/a	n/a	SEQ NO: 8	SEQ NO: 20	n/a	SEQ NO: 30	SEQ NO: 45
Intron #3	n/a	n/a	n/a	n/a	n/a	SEQ NO: 31	SEQ NO: 46
Intron #3A	n/a	n/a	SEQ NO: 9	SEQ NO: 21	n/a	n/a	n/a
Intron #3B	n/a	n/a	SEQ NO: 12	SEQ NO: 22	n/a	n/a	n/a
Intron #3C	n/a	n/a	SEQ NO: 13	SEQ NO: 23	n/a	n/a	n/a
Intron #4	n/a	n/a	SEQ NO: 10	SEQ NO: 24	SEQ NO: 14	SEQ NO: 32	SEQ NO: 47
Intron #5	n/a	n/a	SEQ NO: 11	SEQ NO: 25	n/a	SEQ NO: 33	n/a
Intron #6	n/a	n/a	n/a	n/a	n/a	SEQ NO: 34	n/a
Intron #7	n/a	n/a	n/a	n/a	n/a	SEQ NO: 35	n/a

Example 3 Combination Constructs

- 5 In Figures 7-15, promoters are indicated by arrows, encoding sequences (both coding and non-coding) are indicated by pentagons which point in the direction of transcription, sense sequences are labeled in normal text, and antisense sequences are labeled in upside-down text. The abbreviations used in these Figures include: 7Sa = 7S α promoter; 7Sa' = 7S α ' promoter; Br napin = *Brassica napin*

promoter; FMV = an FMV promoter; ARC = arcelin promoter; RBC E9 3' = Rubisco E9 termination signal; Nos 3' = *nos* termination signal; TML 3' = *tml* termination signal; napin 3' = napin termination signal; '3 (in the same box as FAD or FAT) = 3' UTR; 5' (in the same box as FAD or FAT) = 5'UTR; Cr = *Cuphea pulcherrima*; Gm = Glycine max; Rc = *Ricinus communis*; FAB2 = a FAB2 allele of a stearyl-
5 desaturase gene; and Intr or Int = intron.

3A. *dsRNA Constructs*

FIGS. 7-9 depict nucleic acid molecules of the present invention in which the first sets of DNA sequences are capable of expressing dsRNA constructs. The first set of DNA sequences depicted in FIGS. 7-9 comprise pairs of related sense and antisense sequences, arranged such that, *e.g.*, the RNA
10 expressed by the first sense sequence is capable of forming a double-stranded RNA with the antisense RNA expressed by the first antisense sequence. The sense sequences may be adjacent to the antisense sequences, or separated from the antisense sequences by a spacer sequence, as shown in FIG. 9.

The second set of DNA sequences comprises coding sequences, each of which is a DNA sequence that encodes a sequence that when expressed is capable of increasing one or both of the protein
15 and transcript encoded by a gene selected from the group consisting of *KAS I*, *KAS IV*, delta-9 desaturase, and CP4 *EPSPS*. Each coding sequence is associated with a promoter, which can be any promoter functional in a plant, or any plant promoter, and may be an FMV promoter, a napin promoter, a 7S (either 7S α or 7S α') promoter, an arcelin promoter, a delta-9 desaturase promoter, or a *FAD2-1A* promoter.

Referring now to FIG. 7, soybean *FAD2-1* intron 1 (SEQ ID NO: 1 or 2), *FAD3-1A* 3'UTR (SEQ
20 ID NO: 16), and *FATB-1* 3'UTR (SEQ ID NO: 36) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense and antisense orientations, separated by a spliceable soybean *FAD3-1A* intron 5 (SEQ ID NO: 11), into a vector containing the soybean 7S α' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into
25 pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. Vectors containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica* napin promoter and a *Brassica* napin 3' termination sequence, and a *R. communis* delta-9 desaturase (*FAB2*) gene (SEQ ID NO: 40) regulated by a soybean *FAD2* promoter and

a *nos* 3' termination sequence, are cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, pMON68539, is depicted in FIG. 7 and is used for transformation using methods as described herein.

5 Soybean *FAD2-1* intron 1 (SEQ ID NO: 1 or 2), *FAD3-1A* intron 4 (SEQ ID NO: 10), and *FATB-1* intron II (SEQ ID NO: 30) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense and antisense orientations, separated by a spliceable soybean *FAD3-1A* intron 5 (SEQ ID NO: 11), into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a
10 vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, pMON68540, is depicted in FIG. 7 and is used for transformation using methods as described herein.

15 Soybean *FAD2-1* intron 1 (SEQ ID NO: 1 or 2), *FAD3-1A* intron 4 (SEQ ID NO: 10), and *FATB-1* intron II (SEQ ID NO: 30) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense and antisense orientations, separated by a spliceable soybean *FAD3-1A* intron 5 (SEQ ID NO: 11), into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a
20 vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. A vector containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica* napin promoter and a *Brassica* napin 3' termination sequence is cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, pMON68544, is depicted in FIG. 7 and is used for transformation using methods as described herein.

25 Soybean *FAD2-1* intron 1 (SEQ ID NO: 1 or 2), *FAD3-1A* intron 4 (SEQ ID NO: 10), *FATB-1* intron II (SEQ ID NO: 30), and *FAD3-1B* intron 4 (SEQ ID NO: 24) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense and antisense orientations, separated by a spliceable soybean *FAD3-1A* intron 5 (SEQ ID NO: 11), into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I

and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, pMON68546, is depicted in FIG. 7 and is used for transformation using methods as described herein.

Referring now to FIG. 8, soybean *FAD2-1* intron 1 (SEQ ID NO: 1 or 2), *FAD3-1A* 3'UTR (SEQ ID NO: 16), and *FATB-1* 3'UTR (SEQ ID NO: 36) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense and antisense orientations, separated by a spliceable soybean *FAD3-1A* intron 5 (SEQ ID NO: 11), into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, pMON68536, is depicted in FIG. 8 and is used for transformation using methods as described herein.

Soybean *FAD2-1* intron 1 (SEQ ID NO: 1 or 2), *FAD3-1A* 3'UTR (SEQ ID NO: 16), and *FATB-1* 3'UTR (SEQ ID NO: 36) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense and antisense orientations, separated by a spliceable soybean *FAD3-1A* intron 5 (SEQ ID NO: 11), into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. A vector containing a *R. communis* delta-9 desaturase (*FAB2*) gene (SEQ ID NO: 40) regulated by a soybean *FAD2* promoter and a *nos* 3' termination sequence, is cut with appropriate restriction enzymes, and ligated just upstream of the *tml* 3' termination sequence. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, pMON68537, is depicted in FIG. 8 and is used for transformation using methods as described herein.

Soybean *FAD2-1* intron 1 (SEQ ID NO: 1 or 2), *FAD3-1A* 3'UTR (SEQ ID NO: 16), and *FATB-1* 3'UTR (SEQ ID NO: 36) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense and antisense orientations, separated by a spliceable soybean *FAD3-1A* intron 5 (SEQ ID NO: 11), into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered

onto the 5' ends of the PCR primers. The vector is then cut with *NotI* and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. A vector containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica napin* promoter and a *Brassica napin* 3' termination sequence is cut with appropriate
5 restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, pMON68538, is depicted in FIG. 8 and is used for transformation using methods as described herein.

Referring now to FIG. 9, soybean *FAD2-1* 3'UTR (SEQ ID NO: 5), *FATB-1* 3'UTR (SEQ ID NO: 36), *FAD3-1A* 3'UTR (SEQ ID NO: 16), and *FAD3-1B* 3'UTR (SEQ ID NO: 26) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The
10 PCR products are cloned directly, in sense and antisense orientations, separated by a spliceable soybean *FAD3-1A* intron 5 (SEQ ID NO: 11), into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *XhoI* sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *NotI* and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression
15 construct, pMON80622, is depicted in FIG. 9 and is used for transformation using methods as described herein.

Soybean *FAD2-1* 3'UTR (SEQ ID NO: 5), *FATB-1* 3'UTR (SEQ ID NO: 36), and *FAD3-1A* 3'UTR (SEQ ID NO: 16) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense and antisense
20 orientations, separated by a spliceable soybean *FAD3-1A* intron 5 (SEQ ID NO: 11), into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *XhoI* sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *NotI* and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, pMON80623, is depicted in FIG. 9 and is
25 used for transformation using methods as described herein.

Soybean *FAD2-1* 5'UTR-3'UTR (SEQ ID NOs: 6 and 5, ligated together), *FATB-1* 5'UTR-3'UTR (SEQ ID NOs: 37 and 36, ligated together), *FAD3-1A* 3'UTR (SEQ ID NO: 16) and *FAD3-1B* 5'UTR-3'UTR (SEQ ID NOs: 27 and 26, ligated together) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned

directly, in sense and antisense orientations, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *XhoI* sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *NotI* and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, O5, is depicted in FIG. 9 and is used for transformation using methods as described herein.

Soybean *FAD2-1* 5'UTR-3'UTR (SEQ ID NOS: 6 and 5, ligated together), *FATB-1* 5'UTR-3'UTR (SEQ ID NOS: 37 and 36, ligated together) and *FAD3-1A* 3'UTR (SEQ ID NO: 16) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense and antisense orientations, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *XhoI* sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *NotI* and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. A vector containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica napin* promoter and a *Brassica napin* 3' termination sequence is cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, O6, is depicted in FIG. 9 and is used for transformation using methods as described herein.

3B. Sense Cosuppression Constructs

FIGS. 10-13 and 19-20 depict nucleic acid molecules of the present invention in which the first sets of DNA sequences are capable of expressing sense cosuppression constructs. The second set of DNA sequences comprises coding sequences, each of which is a DNA sequence that encodes a sequence that when expressed is capable of increasing one or both of the protein and transcript encoded by a gene selected from the group consisting of *KAS I*, *KAS IV*, delta-9 desaturase, and CP4 *EPSPS*. Each coding sequence is associated with a promoter, which is any promoter functional in a plant, or any plant promoter, and may be an FMV promoter, a napin promoter, a 7S promoter (either 7S α or 7S α '), an arcelin promoter, a delta-9 desaturase promoter, or a *FAD2-1A* promoter.

Referring now to FIG. 10, soybean *FAD2-1* intron 1 (SEQ ID NO: 1 or 2), *FAD3-1C* intron 4 (SEQ ID NO: 14), *FATB-1* intron II (SEQ ID NO: 30), *FAD3-1A* intron 4 (SEQ ID NO: 10), and *FAD3-*

1B intron 4 (SEQ ID NO: 24) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α ' promoter and a pea Rubisco E9 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, pMON68522, is depicted in FIG. 10 and is used for transformation using methods as described herein.

Soybean *FAD2-1* intron 1 (SEQ ID NO: 1 or 2), *FAD3-1A* intron 4 (SEQ ID NO: 10), *FAD3-1B* intron 4 (SEQ ID NO: 24), and *FATB-1* intron II (SEQ ID NO: 30) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. Vectors containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica napin* promoter and a *Brassica napin* 3' termination sequence, and a *R. communis* delta-9 desaturase (*FAB2*) gene (SEQ ID NO: 40) regulated by a soybean *FAD2* promoter and a *nos* 3' termination sequence, are cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, pMON80614, is depicted in FIG. 10 and is used for transformation using methods as described herein.

Soybean *FAD2-1* intron 1 (SEQ ID NO: 1 or 2), *FAD3-1A* 3'UTR (SEQ ID NO: 16), and *FATB-1* 3'UTR (SEQ ID NO: 36) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, pMON68531, is depicted in FIG. 10 and is used for transformation using methods as described herein.

Soybean *FAD2-1* intron 1 (SEQ ID NO: 1 or 2), *FAD3-1A* 3'UTR (SEQ ID NO: 16), and *FATB-1* 3'UTR (SEQ ID NO: 36) sequences are amplified via PCR to result in PCR products that include

reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. Vectors containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica napin* promoter and a *Brassica napin* 3' termination sequence, and a *R. communis* delta-9 desaturase (*FAB2*) gene (SEQ ID NO: 40) regulated by a soybean *FAD2* promoter and a *nos* 3' termination sequence, are cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, pMON68534, is depicted in FIG. 10 and is used for transformation using methods as described herein.

Soybean *FAD2-1* intron 1 (SEQ ID NO: 1 or 2), *FAD3-1A* 3'UTR (SEQ ID NO: 16), and *FATB-1* 3'UTR (SEQ ID NO: 36) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. A vector containing a *R. communis* delta-9 desaturase (*FAB2*) gene (SEQ ID NO: 40) regulated by a soybean *FAD2* promoter and a *nos* 3' termination sequence, is cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, pMON68535, is depicted in FIG. 10 and is used for transformation using methods as described herein.

Referring now to FIG. 11, soybean *FAD2-1* 3'UTR (SEQ ID NO: 5), *FAD3-1A* 3'UTR (SEQ ID NO: 16), and *FATB-1* 3'UTR (SEQ ID NO: 36) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, pMON80605, is depicted in FIG. 11 and is used for transformation using methods as described herein.

Soybean *FAD2-1* 3'UTR (SEQ ID NO: 5), *FAD3-1A* 3'UTR (SEQ ID NO: 16), and *FATB-1* 3'UTR (SEQ ID NO: 36) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. A vector containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica napin* promoter and a *Brassica napin* 3' termination sequence is cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, pMON80606, is depicted in FIG. 11 and is used for transformation using methods as described herein.

Soybean *FAD2-1* 3'UTR (SEQ ID NO: 5), *FAD3-1A* 3'UTR (SEQ ID NO: 16), and *FATB-1* 3'UTR (SEQ ID NO: 36) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. A vector containing a *R. communis* delta-9 desaturase (*FAB2*) gene (SEQ ID NO: 40) regulated by a soybean *FAD2* promoter and a *nos* 3' termination sequence is cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, pMON80607, is depicted in FIG. 11 and is used for transformation using methods as described herein.

Soybean *FAD2-1* 3'UTR (SEQ ID NO: 5), *FAD3-1A* 3'UTR (SEQ ID NO: 16), and *FATB-1* 3'UTR (SEQ ID NO: 36) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. Vectors containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica napin* promoter and a *Brassica napin* 3' termination sequence, and a *R. communis* delta-9 desaturase (*FAB2*) gene (SEQ ID NO: 40) regulated by a soybean *FAD2* promoter and

a *nos* 3' termination sequence, are cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, pMON80614, is depicted in FIG. 11 and is used for transformation using methods as described herein.

Referring now to FIG. 12, soybean *FAD2-1* 3'UTR (SEQ ID NO: 5), *FATB-1* 3'UTR (SEQ ID NO: 36), and *FAD3-1A* 3'UTR (SEQ ID NO: 16) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, pMON80629, is depicted in FIG. 12 and is used for transformation using methods as described herein.

Soybean *FAD2-1* intron 1 (SEQ ID NO: 1 or 2), *FAD3-1A* intron 4 (SEQ ID NO: 10), *FATB-1* intron II (SEQ ID NO: 30), and *FAD3-1A* intron 4 (SEQ ID NO: 10) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, pMON81902, is depicted in FIG. 12 and is used for transformation using methods as described herein.

Soybean *FAD2-1* 5'UTR-3'UTR (SEQ ID NOS: 6 and 5, ligated together), *FAD3-1* 5'UTR-3'UTR (SEQ ID NOS: 17 and 16, ligated together, or 27 and 26, ligated together), and *FATB-1* 5'UTR-3'UTR (SEQ ID NOS: 37 and 36, ligated together) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The *FAD2-1* PCR product is cloned directly, in sense orientation, into a vector containing the soybean 7S α promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. Similarly, the *FAD3-1* PCR product is cloned directly, in sense orientation, into a vector containing the soybean 7S α promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The *FATB-1* PCR product is cloned directly, in sense orientation, into a vector containing the arcelin

promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. These vectors are then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, O1, is depicted in FIG. 12 and is used for transformation using methods as described herein.

Soybean *FAD2-1* 5'UTR-3'UTR (SEQ ID NOs: 6 and 5, ligated together), *FAD3-1* 5'UTR-3'UTR (SEQ ID NOs: 17 and 16, ligated together, or 27 and 26, ligated together), and *FATB-1* 5'UTR-3'UTR (SEQ ID NOs: 37 and 36, ligated together) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The *FAD2-1* PCR product is cloned directly, in sense orientation, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. Similarly, the *FAD3-1* PCR product is cloned directly, in sense orientation, into a vector containing the soybean 7S α promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The *FATB-1* PCR product is cloned directly, in sense orientation, into a vector containing the arcelin promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. These vectors are then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. A vector containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica napin* promoter and a *Brassica napin* 3' termination sequence is cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, O2, is depicted in FIG. 12 and is used for transformation using methods as described herein.

Referring now to FIG. 13, soybean *FAD2-1* 5'UTR-3'UTR (SEQ ID NOs: 6 and 5, ligated together), *FATB-1* 5'UTR-3'UTR (SEQ ID NOs: 37 and 36, ligated together), *FAD3-1A* 3'UTR (SEQ ID NO: 16), and *FAD3-1B* 5'UTR-3'UTR (SEQ ID NOs: 27 and 26, ligated together) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vectors are then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. A vector

containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica napin* promoter and a *Brassica napin* 3' termination sequence is cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, O7, is depicted in FIG. 13 and is used for transformation using methods as described herein.

5. Soybean *FAD2-1* intron 1 (SEQ ID NO: 1 or 2) is amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. Soybean *FATB-1* 5'UTR-3'UTR (SEQ ID NOs: 37 and 36, ligated together), *FAD3-1A* 3'UTR (SEQ ID NO: 16), and *FAD3-1B* 5'UTR-3'UTR (SEQ ID NOs: 27 and 26, ligated together) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α promoter and a *nos* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vectors are then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. A vector containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica napin* promoter and a *Brassica napin* 3' termination sequence is cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, O9, is depicted in FIG. 13 and is used for transformation using methods as described herein.

Referring now to FIG. 19, soybean *FATB-2* non-coding sequences (SEQ ID NOs: 44-47), *FAD2-1* non-coding sequences (SEQ ID NOs: 1 and 5-6), and *FATB-1* non-coding sequences (SEQ ID NOs: 29-37) are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vectors are then cut with *Not*I and ligated into pMON80612, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct is depicted in FIG. 19-A and is used for transformation using methods described herein.

A DNA sequence containing a delta-9 desaturase is regulated by a 7S alpha promoter and a *TML* 3' termination sequence is cut using the appropriate restriction enzymes and ligated into the above

expression construct. The resulting expression construct is depicted in FIG. 19-B and is used for transformation using methods described herein.

5 A vector containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a bean arcelin promoter and a napin 3' termination sequence is cut with appropriate restriction enzymes, and ligated into the above expression construct. The resulting gene expression construct is depicted in FIG. 19-C and is used for transformation using methods as described herein.

10 Referring now to FIG. 20 soybean *FATB-2* non-coding sequences (SEQ ID NOs: 44-47), *FAD2-1* non-coding sequences (SEQ ID NOs: 1 and 5-6), *FATB-1* non-coding sequences (SEQ ID NOs: 29-37), *FAD3-1A* non-coding sequences (SEQ ID NOs: 7-13 and 16-17), and *FAD3-1B* non-coding sequences (SEQ ID NOs: 19-27) are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in sense orientation, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vectors are then cut with *Not*I and ligated into pMON80612, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct is depicted in FIG. 20-A and is used for transformation using methods described herein.

15 A DNA sequence containing a delta-9 desaturase is regulated by a 7S alpha promoter and a *TML* 3' termination sequence is cut using the appropriate restriction enzymes and ligated into the above expression construct. The resulting expression construct is depicted in FIG. 20-B and is used for transformation using methods described herein.

20 A vector containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica* bean arcelin promoter and a napin 3' termination sequence is cut with appropriate restriction enzymes, and ligated into the above expression construct. The resulting gene expression construct is depicted in FIG. 20-C and is used for transformation using methods as described herein.

25 3C. Antisense Constructs

FIG. 14 depicts nucleic acid molecules of the present invention in which the first sets of DNA sequences are capable of expressing antisense constructs, and FIGs. 15 through 18 depict nucleic acid molecules of the present invention in which the first sets of DNA sequences are capable of expressing

combinations of sense and antisense constructs. The second set of DNA sequences comprises coding sequences, each of which is a DNA sequence that encodes a sequence that when expressed is capable of increasing one or both of the protein and transcript encoded by a gene selected from the group consisting of *KAS I*, *KAS IV*, delta-9 desaturase, and CP4 *EPSPS*. Each coding sequence is associated with a promoter, which is any promoter functional in a plant; or any plant promoter, and may be an FMV promoter, a napin promoter, a 7S (either 7S α or 7S α') promoter, an arcelin promoter, a delta-9 desaturase promoter, or a *FAD2-1A* promoter.

Referring now to FIG. 14, soybean *FAD2-1* 3'UTR (SEQ ID NO: 5), *FATB-1* 3'UTR (SEQ ID NO: 36), and *FAD3-1A* 3'UTR (SEQ ID NO: 16) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in antisense orientation, into a vector containing the soybean 7S α' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, pMON80615, is depicted in FIG. 14 and is used for transformation using methods as described herein.

Soybean *FAD2-1* 3'UTR (SEQ ID NO: 5), *FATB-1* 3'UTR (SEQ ID NO: 36), and *FAD3-1A* 3'UTR (SEQ ID NO: 16) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in antisense orientation, into a vector containing the soybean 7S α' promoter and a *tml* 3' termination sequence, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. A vector containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica* napin promoter and a *Brassica* napin 3' termination sequence is cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, pMON80616, is depicted in FIG. 14 and is used for transformation using methods as described herein.

Soybean *FAD2-1* 3'UTR (SEQ ID NO: 5), *FATB-1* 3'UTR (SEQ ID NO: 36), and *FAD3-1A* 3'UTR (SEQ ID NO: 16) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in antisense orientation, into a vector containing the soybean 7S α' promoter and a *tml* 3' termination sequence, by way of *Xho*I

sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *NotI* and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. A vector containing a *R. communis* delta-9 desaturase (*FAB2*) gene (SEQ ID NO: 40) regulated by a soybean *FAD2* promoter and a *nos* 3' termination sequence, is cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, pMON80617, is depicted in FIG. 14 and is used for transformation using methods as described herein.

Soybean *FAD2-1* 3'UTR (SEQ ID NO: 5), *FATB-1* 3'UTR (SEQ ID NO: 36), and *FAD3-1A* 3'UTR (SEQ ID NO: 16) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in antisense orientation, into a vector containing the soybean 7S α promoter and a *tml* 3' termination sequence, by way of *XhoI* sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *NotI* and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, pMON80630, is depicted in FIG. 14 and is used for transformation using methods as described herein.

Soybean *FAD2-1* 5'UTR-3'UTR (SEQ ID NOs: 6 and 5, ligated together), *FATB-1* 5'UTR-3'UTR (SEQ ID NOs: 37 and 36, ligated together), *FAD3-1A* 3'UTR (SEQ ID NO: 16), and *FAD3-1B* 5'UTR-3'UTR (SEQ ID NOs: 27 and 26, ligated together) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly, in antisense orientation, into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, by way of *XhoI* sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *NotI* and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. A vector containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica napin* promoter and a *Brassica napin* 3' termination sequence is cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, O8, is depicted in FIG. 14 and is used for transformation using methods as described herein.

Referring now to FIG. 15, soybean *FAD2-1* 5'UTR-3'UTR (SEQ ID NOs: 6 and 5, ligated together), *FAD3-1A* 5'UTR-3'UTR (SEQ ID NOs: 17 and 16, ligated together), and *FATB-1* 5'UTR-3'UTR (SEQ ID NOs: 37 and 36, ligated together) sequences are amplified via PCR to result in PCR

products that include reengineered restriction sites at both ends. The PCR products are cloned directly in sense and antisense orientation into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, with an additional soybean 7S α promoter located between the sense and antisense sequences, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct, O3, is depicted in FIG. 15 and is used for transformation using methods as described herein.

Soybean *FAD2-1* 5'UTR-3'UTR (SEQ ID NOs: 6 and 5, ligated together), *FAD3-1A* 5'UTR-3'UTR (SEQ ID NOs: 27 and 26, ligated together), and *FATB-1* 5'UTR-3'UTR (SEQ ID NOs: 37 and 36, ligated together) sequences are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly in sense and antisense orientation into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence, with an additional soybean 7S α promoter located between the sense and antisense sequences, by way of *Xho*I sites engineered onto the 5' ends of the PCR primers. The vector is then cut with *Not*I and ligated into pMON41164, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. A vector containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a *Brassica napin* promoter and a *Brassica napin* 3' termination sequence is cut with appropriate restriction enzymes, and ligated into pMON41164. The resulting gene expression construct, O4, is depicted in FIG. 15 and is used for transformation using methods as described herein.

Referring now to FIG. 16, soybean *FATB-2* non-coding sequences (SEQ ID NOs: 44-47), *FATB-1* non-coding sequences (SEQ ID NOs: 29-37), and *FAD2-1* non-coding sequences (SEQ ID NOs: 1 and 5-6) are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly in sense and antisense orientation into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence. The vector is then cut with with an appropriate restriction endonuclease and ligated into pMON80612 a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct is depicted in FIG. 16-A and is used for transformation using methods as described herein.

A DNA sequence containing a delta-9 desaturase is regulated by a 7S alpha promoter and a *TML* 3' termination sequence is cut using the appropriate restriction enzymes and ligated into the above expression construct. The resulting expression construct is depicted in FIG. 16-B and is used for transformation using methods described herein.

5 A vector containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a bean arcelin promoter and a napin 3' termination sequence is cut with appropriate restriction enzymes, and ligated into the above expression construct. The resulting gene expression construct is depicted in FIG. 16-C and is used for transformation using methods as described herein.

10 Referring now to FIG. 17, soybean *FATB-2* non-coding sequences (SEQ ID NOs: 44-47), *FATB-1* non-coding sequences (SEQ ID NOs: 29-37), *FAD2-1* non-coding sequences (SEQ ID NOs: 1 and 5-6), and *FAD3-1A* non-coding sequences (SEQ ID NOs: 7-13 and 16-17) are amplified via PCR to result in PCR products that include reengineered restriction sites at both ends. The PCR products are cloned directly in sense and antisense orientation into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence. The vector is then cut with with an appropriate restriction endonuclease and
15 ligated into pMON80612, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The resulting gene expression construct is depicted in FIG. 17-A and is used for transformation using methods as described herein.

A DNA sequence containing a delta-9 desaturase is regulated by a 7S alpha promoter and a *TML* 3' termination sequence is cut using the appropriate restriction enzymes and ligated into the above
20 expression construct. The resulting expression construct is depicted in FIG. 17-B and is used for transformation using methods described herein.

A vector containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a bean arcelin promoter and a napin 3' termination sequence is cut with appropriate restriction enzymes, and ligated into the above expression construct. The resulting gene expression construct is depicted in FIG. 17-C and is
25 used for transformation using methods as described herein.

Referring now to FIG. 18, soybean *FATB-2* non-coding sequences (SEQ ID NOs: 44-47), *FATB-1* non-coding sequences (SEQ ID NOs: 29-37), *FAD2-1* non-coding sequences (SEQ ID NOs: 1 and 5-6), *FAD3-1A* non-coding sequences (SEQ ID NOs: 7-13 and 16-17) and *FAD3-1B* non-coding sequences (SEQ ID NOs: 19-27) are amplified via PCR to result in PCR products that include reengineered

restriction sites at both ends. The PCR products are cloned directly in sense and antisense orientation into a vector containing the soybean 7S α ' promoter and a *tml* 3' termination sequence. The vector is then cut with with an appropriate restriction endonuclease and ligated into pMON80612, a vector that contains the CP4 *EPSPS* gene regulated by the FMV promoter and a pea Rubisco E9 3' termination sequence. The
5 resulting gene expression construct is depicted in FIG. 18-A and is used for transformation using methods as described herein.

A DNA sequence containing a delta-9 desaturase is regulated by a 7S alpha promoter and a *TML* 3' termination sequence is cut using the appropriate restriction enzymes and ligated into the above expression construct. The resulting expression construct is depicted in FIG. 18-B and is used for
10 transformation using methods described herein.

A vector containing a *C. pulcherrima* KAS IV gene (SEQ ID NO: 39) regulated by a bean arcelin promoter and a napin 3' termination sequence is cut with appropriate restriction enzymes, and ligated into the above expression construct. The resulting gene expression construct is depicted in FIG. 18-C and is used for transformation using methods as described herein. The above-described nucleic acid molecules
15 are preferred embodiments which achieve the objects, features and advantages of the present invention. It is not intended that the present invention be limited to the illustrated embodiments. The arrangement of the sequences in the first and second sets of DNA sequences within the nucleic acid molecule is not limited to the illustrated and described arrangements, and may be altered in any manner suitable for achieving the objects, features and advantages of the present invention as described herein, illustrated in
20 the accompanying drawings, and encompassed within the claims.

Example 4 Plant Transformation and Analysis

The constructs of Examples 2 and 3 are stably introduced into soybean (for example, Asgrow variety A4922 or Asgrow variety A3244 or Asgrow variety A3525) by the methods described earlier, including the methods of McCabe *et al.*, *Bio/Technology*, 6:923-926 (1988), or *Agrobacterium*-mediated
25 transformation. Transformed soybean plants are identified by selection on media containing glyphosate. Fatty acid compositions are analyzed from seed of soybean lines transformed with the constructs using gas chromatography. In addition, any of the constructs may contain other sequences of interest, as well as different combinations of promoters.

For some applications, modified fatty acid compositions are detected in developing seeds, whereas in other instances, such as for analysis of oil profile, detection of fatty acid modifications occurring later in the FAS pathway, or for detection of minor modifications to the fatty acid composition, analysis of fatty acid or oil from mature seeds is preferred. Furthermore, analysis of oil and/or fatty acid content of individual seeds may be desirable, especially in detection of oil modification in the segregating R1 seed populations. As used herein, R0 indicates the plant and seed arising from transformation/regeneration protocols described herein, and R1 indicates plants and seeds generated from the transgenic R0 seed.

Fatty acid compositions are determined for the seed of soybean lines transformed with the constructs of Example 3. One to ten seeds of each of the transgenic and control soybean lines are ground individually using a tissue homogenizer (Pro Scientific) for oil extraction. Oil from ground soybean seed is extracted overnight in 1.5 ml heptane containing triheptadecanoin (0.50 mg/ml). Aliquots of 200 μ l of the extracted oil are derivatized to methyl esters with the addition of 500 μ l sodium methoxide in absolute methanol. The derivatization reaction is allowed to progress for 20 minutes at 50°C. The reaction is stopped by the simultaneous addition of 500 μ l 10% (w/v) sodium chloride and 400 μ l heptane. The resulting fatty acid methyl esters extracted in hexane are resolved by gas chromatography (GC) on a Hewlett-Packard model 6890 GC (Palo Alto, CA). The GC was fitted with a Supelcowax 250 column (30 m, 0.25 mm id, 0.25 micron film thickness) (Supelco, Bellefonte, PA). Column temperature is 175°C at injection and the temperature programmed from 175°C to 245°C to 175°C at 40°C/min. Injector and detector temperatures are 250°C and 270°C, respectively.

Example 5 Synthesized Fuel Oil with Improved Biodiesel Properties

A synthesized fuel oil fatty acid composition is prepared having the following mixtures of fatty acid methyl esters: 73.3% oleic acid, 21.4% linoleic acid, 2.2% palmitic acid, 2.1% linolenic acid and 1.0% stearic acid (all by weight). Purified fatty acid methyl esters are obtained from Nu-Chek Prep, Inc., Elysian, MN, USA. The cetane number and ignition delay of this composition is determined by the Southwest Research Institute using an Ignition Quality Tester ("IQT") 613 (Southwest Research Institute, San Antonio, Texas, USA).

An IQT consists of a constant volume combustion chamber that is electrically heated, a fuel injection system, and a computer that is used to control the experiment, record the data and provide interpretation of the data. The fuel injection system includes a fuel injector nozzle that forms an entrance to the combustion chamber. A needle lift sensor in the fuel injector nozzle detects fuel flow into the combustion chamber. A pressure transducer attached to the combustion chamber measures cylinder pressure, the pressure within the combustion chamber. The basic concept of an IQT is measurement of the time from the start of fuel injection into the combustion chamber to the start of combustion. The thermodynamic conditions in the combustion chamber are precisely controlled to provide consistent measurement of the ignition delay time.

For a cetane number and ignition delay test, the test fuel is filtered using a 5-micron filter. The fuel reservoir, injection line, and nozzle are purged with pressurized nitrogen. The fuel reservoir is then cleaned with a lint free cloth. A portion of the test fuel is used to flush the fuel reservoir, injection line, and nozzle. The reservoir is filled with the test fuel and all air is bled from the system. The reservoir is pressurized to 50 psig. The method basically consists of injecting at high pressure a precisely metered quantity of the test fuel into the combustion chamber that is charged with air to the desired pressure and temperature. The measurement consists of determining the time from the start of injection to the onset of combustion, often referred to as the ignition delay time. This determination is based on the measured needle lift and combustion chamber pressures. The normal cetane rating procedure calls for setting the skin temperature at 567.5°C and the air pressure at 2.1 MPa.

A fuel with a known injection delay is run in the IQT combustion bomb at the beginning of the day to make sure the unit is operating within normal parameters. The test synthetic is then run. The known fuel is run again to verify that the system has not changed. Once the fuel reservoir is reconnected to the fuel injection pump, the test procedure is initiated on the PC controller. The computer controls all the procedure, including the air charging, fuel injection, and exhaust events. 32 repeat combustion events are undertaken.

The ignition delay is the time from the start of injection to the start of ignition. It is determined from the needle lift and cylinder pressure data. The rise of the injection needle signals start of injection. The cylinder pressure drops slightly due to the cooling effect of the vaporization of the fuel. Start of

combustion is defined as the recovery time of the cylinder pressure – increases due to combustion to the pressure it was just prior to fuel injection.

The measured ignition delay times are then used to determine the cetane number based on a calibration curve that is incorporated into the data acquisition and reduction software. The calibration curve, consisting of cetane number as a function of ignition delay time, is generated using blends of the primary reference fuels and NEG check fuels. In the case of test fuels that are liquid at ambient conditions, the calibration curve is checked on a daily basis using at least one check fuel of known cetane number (Ryan, “Correlation of Physical and Chemical Ignition Delay to Cetane Number”, SAE Paper 852103 (1985); Ryan, “Diesel Fuel Ignition Quality as Determined in a Constant Volume Combustion Bomb”, SAE Paper 870586 (1986); Ryan, “Development of a Portable Fuel Cetane Quality Monitor”, Belvoir Fuels and Lubricants Research Facility Report No. 277, May (1992); Ryan, “Engine and Constant Volume Bomb Studies of Diesel Ignition and Combustion”, SAE Paper 881616 (1988); and Allard *et al.*, “Diesel Fuel Ignition Quality as Determined in the Ignition Quality Tester (“IQT”)”, SAE Paper 961182 (1996)). As shown in Table 3, the synthesized oil composition exhibits cetane numbers and ignition delays that are suitable for use for example, without limitation, as a biodiesel oil.

TABLE 3

Fuel Name	Test Number	Cetane Number	Std.Dev. Cetane No.	Ignition Delay (ms)	Std.Dev. Ign. Delay
Check-High ¹	1777	49.55	0.534	4.009	0.044
Check-High	1778	49.33	0.611	4.028	0.051
	Average	49.4		4.02	
Synthesized Oil	1779	55.02	1.897	3.622	0.116
Synthesized Oil	1780	55.65	1.807	3.583	0.109
Synthesized Oil	1781	55.63	1.649	3.583	0.098
	Average	55.4		3.60	
Check-High	1786	49.2	0.727	4.04	0.061

¹ The fuel called "Check-High" is a calibration fuel. It should have a cetane number of 49.3 ± 0.5 . The unit is checked with the calibration before and after running the synthetic test fuel.

5 The density (ASTM D-4052) cloud point (ASTM D-2500), pour point (ASTM D-97), and cold filter plugging point (IP 309/ASTM D-6371) are determined for the synthesized oil using ASTM D protocols. ASTM D protocols are obtained from ASTM, 100 Barr Harbor Drive, West Conshohocken, PA, USA. The results of these tests are set forth in Table 4. As shown in Table 4, the synthesized oil composition exhibits numbers that are suitable for use as, for example without limitation, as a biodiesel oil.

10

TABLE 4

TEST	METHOD	RESULTS
Density	ASTM D-4052	0.8791 g/mL
Cloud Point	ASTM D-2500	-18 deg. C
Pour Point	ASTM D-97	-21 deg. C
Cold Filter Plugging Point	IP 309 (same as ASTM D-6371)	-21 deg. C

15 Levels of nitric oxide emissions are estimated by evaluating the unsaturation levels of a biologically-based fuel, by measuring the fuel density and indirectly calculating the estimated emissions levels, or by directly measuring. There are also standard protocols available for directly measuring levels of nitric oxide emissions. The synthesized oil is estimated to have lower nitric oxide emissions levels than methyl esters of fatty acids made from conventional soybean oil based on an evaluation of the overall level of unsaturation in the synthesized oil. Oils containing larger numbers of double bonds, *i.e.*, having a higher degree of unsaturation, tend to produce higher nitric oxide emissions. The oil has a total

of 123 double bonds, as compared to conventional soybean oil's total of 153 double bonds, as shown in Table 5.

TABLE 5

SYNTHETIC OIL		
73 % oleic acid (18:1)	x 1 double bond =	73
22 % linoleic acid (18:2)	x 2 double bonds =	44
2 % linolenic acid (18:3)	x 3 double bonds =	6
TOTAL double bonds		123
CONVENTIONAL SOYBEAN OIL		
23 % oleic acid (18:1)	x 1 double bond =	23
53 % linoleic acid (18:2)	x 2 double bonds =	106
8 % linolenic acid (18:3)	x 3 double bonds =	24
TOTAL double bonds		153

5 As reported by the National Renewable Energy Laboratory, Contract No. ACG-8-17106-02 Final Report, *The Effect Of Biodiesel Composition On Engine Emissions From A DDC Series 60 Diesel Engine*, (June 2000), nitric acid emissions of biodiesel compositions are predicted by the formula $y = 46.959x - 36.388$ where y is the oxide emissions in grams/brake horse power hours; and x is the density of biodiesel. The formula is based on a regression analysis of nitric acid emission data in a test involving 16
10 biodiesel fuels. The test makes use of a 1991 calibration, production series 60 model Detroit Diesel Corporation engine.

The density of the synthesized oil is determined by Southwest Research Institute using the method ASTM D4052. The result shown in Table 4 is used in the above equation to predict a nitric oxide emission value of 4.89 g/bhp-h. This result is compared to a control soybean product. The National
15 Renewable Energy Laboratory report gives the density and nitric oxide emissions of a control soy based biodiesel (methyl soy ester IGT). The density of the control biodiesel is 0.8877 g/mL, giving a calculated nitric oxide emission of 5.30 g/bhp-h. This calculated emission value is similar to the experimental value for nitric oxide emission of 5.32 g/bhp-h. The synthesized oil composition exhibits improved numbers compared to the control and is suitable for use, for example without limitation, as a biodiesel oil.

Example 6 Optimum Fatty Acid Composition For Healthy Serum Lipid Levels

The cholesterol lowering properties of vegetable compositions are determined to identify fatty acid compositions that have a more favorable effect on serum lipid levels than conventional soybean oil (i.e., lower LDL-cholesterol and higher HDL-cholesterol). Published equations based on 27 clinical trials (Mensink, R.P. and Katan, M.B. *Arteriosclerosis and Thrombosis*, 12:911-919 (1992)) are used to compare the effects on serum lipid levels in humans of new oilseed compositions with that of normal soybean oil.

Table 6 below presents the results of the change in serum lipid levels where 30% of dietary energy from carbohydrate is substituted by lipids. The results show that soybean oil already has favorable effects on serum lipids when it replaces carbohydrates in the diet. Improvements on this composition are possible by lowering saturated fat levels and by obtaining a linoleic acid level between 10-30% of the total fatty acids, preferably about 15-25% of the total fatty acids. When the proportion of linoleic acid is less than 10% of the total fatty acids, the new composition raises LDL-cholesterol compared to control soybean oil, even though the saturated fat content is lowered to 5% of the total fatty acids. When the proportion of linoleic acid is increased, the ability of the composition to raise serum HDL levels is reduced. Therefore, the preferred linoleic acid composition is determined to be about 15-25% of the total fatty acids.

Table 6

	Fatty acids						Serum Lipids
	C16:0	C18:0	C18:1	C18:2	C18:3	Other (C20:1)	
Soy control (%)	11.000	4.000	23.400	53.200	7.800	0.600	
Proportion of 30% fat E (%)	3.300	1.200	7.020	15.960	2.340	0.180	
LDL Calculation (mg/dl)	4.224	1.536	1.685	8.778	1.287	0.043	-6.033
HDL Calc (mg/dl)	1.551	0.564	2.387	4.469	0.655	0.061	9.687
3% 18:2, <6% sat (%)	3.000	2.000	85.000	3.000	3.000	4.000	
Proportion of 30% fat E (%)	0.900	0.600	25.500	0.900	0.900	1.200	
LDL Calculation (mg/dl)	1.152	0.768	6.120	0.495	0.495	0.288	-5.478
vs. control (mg/dl)							0.555
HDL calculation (mg/dl)	0.423	0.282	8.670	0.252	0.252	0.408	10.287
vs. control (mg/dl)							0.600

	Fatty acids						Serum Lipids
	C16:0	C18:0	C18:1	C18:2	C18:3	Other (C20:1)	
10% 18:2, <6% sat (%)	3.000	2.000	72.000	10.000	3.000	10.000	
Proportion of 30% fat E (%)	0.900	0.600	21.600	3.000	0.900	3.000	
LDL Calculation (mg/dl)	1.152	0.768	5.184	1.650	0.495	0.720	-6.129
vs. control (mg/dl)							-0.096
HDL calculation (mg/dl)	0.423	0.282	7.344	0.840	0.252	1.020	10.161
vs. control (mg/dl)							0.474
20% 18:2, <6% sat (%)	3.000	2.000	65.000	20.000	3.000	7.000	
Proportion of 30% fat E (%)	0.900	0.600	19.500	6.000	0.900	2.100	
LDL Calculation (mg/dl)	1.152	0.768	4.680	3.300	0.495	0.504	-7.059
vs. control (mg/dl)							-1.026
HDL calculation (mg/dl)	0.423	0.282	6.630	1.680	0.252	0.714	9.981
vs. control (mg/dl)							0.294
21% 18:2, <3.2% sat (%)	2.000	1.000	72.000	21.000	1.000	3.000	
Proportion of 30% fat E (%)	0.600	0.300	21.600	6.300	0.300	0.900	
LDL Calculation (mg/dl)	0.768	0.384	5.184	3.465	0.165	0.216	-7.878
vs. control (mg/dl)							-1.845
HDL calculation (mg/dl)	0.282	0.141	7.344	1.764	0.084	0.306	9.921
vs. control (mg/dl)							0.234
30% 18:2, <6% sat (%)	3.000	2.000	57.000	30.000	3.000	5.000	
Proportion of 30% fat E (%)	0.900	0.600	17.100	9.000	0.900	1.500	
LDL Calculation (mg/dl)	1.152	0.768	4.104	4.950	0.495	0.360	-7.989
vs. control (mg/dl)							-1.956
HDL calculation (mg/dl)	0.423	0.282	5.814	2.520	0.252	0.510	9.801
vs. control (mg/dl)							0.114

Example 7

The following fourteen steps illustrate the construction of vector pMON68537 designed for plant transformation to suppress *FAD2*, *FAD3*, and *FATB* genes and overexpress delta-9 desaturase in soybean. In particular, the construct comprises a 7S alpha promoter operably linked to soybean sense-oriented intron and 3'UTRs, i.e., a *FAD2-1A* intron #1, a *FAD3-1A* 3'UTR, a *FATB-1* 3'UTR, a hairpin loop-forming spliceable intron, and a complementary series of soybean anti-sense-oriented intron and 3'UTR's, i.e., a *FATB-1* 3'UTR, a *FAD3-1A* 3'UTR and a *FAD2-1A* intron #1 and the soybean *FAD2* promoter driving the delta-9 desaturase.

Step1 – The soybean *FAD3-1A* intron #5, which serves as the spliceable intron portion of the RNAi construct, is PCR amplified using soybean genomic DNA as template, with the following primers:

5' primer = 19037 = ACTAGTATATTGAGCTCATATTCCACTGCAGTGGATATT
GTTTAAACATAGCTAGCATATTACGCGTATATTATAACAAGCTTATATTCCCGGGATA
TTGTCGACATATTAGCGGTACATTTTATTGCTTATTAC

3' primer = 19045 = ACTAGTATATTGAGCTCATATTCCTGCAGGATATTCTCGAG
5 ATATTCACGGTAGTAATCTCCAAGAACTGGTTTTGCTGCTTGTGTCTGCAGTGAATC.

These primers add cloning sites to the 5' and 3' ends. To 5' end: SpeI, SacI, BstXI, PmeI, NheI, MluI, HindIII, XmaI, SmaI, Sall. To 3' end: SpeI, SacI, Sse8387I, XhoI. The soybean *FAD3-1A* intron #5 PCR product is cloned into pCR2.1, resulting in KAWHIT03.0065.

KAWHIT03.0065 is then digested with SpeI and the ends are filled with Pfu polymerase and
10 pMON68526 (empty chloramphenicol (hereinafter CM) resistant vector) is digested with HindIII and the ends are filled with Pfu polymerase. KAWHIT03.0065 and pMON68526 are then ligated to create pMON68541 (soybean *FAD3-1A* intron #5 with multiple cloning sites in Amp resistant vector).

Step 2 – The soybean *FATB-1* 3'UTR is amplified with the following primers: 18662=
15 TTTTAATTACAATGAGAATGAGATTTACTGC (adding Bsp120I to the 5' end) and 18661=
GGGCCCGATTTGAAATGGTTAACG. The PCR product is then ligated into pCR2.1 to make KAWHIT03.0036.

Step 3 – KAWHIT03.0036 is then digested with Bsp120I and EcoRI and then cloned into
KAWHIT03.0032 (empty CM resistant vector with a multiple cloning site) to make
20 KAWHIT03.0037 (*FATB-1* 3'UTR in empty CM resistant vector).

Step 4 – The soybean *FAD3-1A* 3'UTR is amplified with the following primers: 18639=
GGGCCCGTTTCAAACCTTTTGG (adding Bsp120I to the 5' end) and 18549=
TGAAACTGACAATTCAA. The PCR product is then ligated into pCR2.1 to make
KAWHIT03.0034.

25 Step 5 – KAWHIT03.0034 is digested with Bsp120I and EcoRI and then ligated into KAWHIT03.0032 (empty CM resistant vector with a multiple cloning site) to make KAWHIT03.0035 (*FAD3-1A* 3'UTR in empty CM resistant vector).

Step 6 – The soybean *FAD 2-1A* intron #1 is PCR amplified using soybean genomic DNA as template, with the following primers: 5' primer = 18663 =

GGGCCCCGGTAAATTAAATTGTGC (Adding Bsp120I site to 5' end); and 3' primer = 18664 = CTGTGTCAAAGTATAACAAGTTCAG. The resulting PCR product is cloned into pCR 2.1 creating KAWHIT03.0038.

5 Step 7 – Soybean *FAD 2-1A* intron #1 PCR product in KAWHIT03.0038 is cloned into KAWHIT03.0032 (empty CM resistant vector with a multiple cloning site) using the restriction sites Bsp120I and EcoRI. The resulting plasmid is KAWHIT03.0039 (soybean *FAD 2-1A* intron #1 in empty CM resistant vector).

10 Step 8 – KAWHIT03.0039 is digested with AscI and HindIII and pMON68541 (*FAD3-1A* intron #5 RNAi AMP resistant base vector) is digested with MluI and HindIII. The soybean *FAD 2-1A* intron #1 is then directionally cloned into pMON68541 to generate KAWHIT03.0071 (soybean *FAD2-1A* intron #1 with soybean *FAD3-1A* intron #5).

15 Step 9 – KAWHIT03.0035 (*FAD3-1A* 3'UTR in CM resistant vector) is digested with AscI and HindIII and KAWHIT03.0071 (*FAD2-1A* intron and *FAD3-1A* intron #5 RNAi AMP resistant base vector) is digested with MluI and HindIII. The soybean *FAD 3-1A* 3'UTR is then directionally cloned into KAWHIT03.0071 to generate KAWHIT03.0072 (soybean *FAD2-1A* intron #1 and *FAD3-1A* 3'UTR with soybean *FAD3-1A* intron #5).

20 Step 10 – KAWHIT03.0037 (*FATB-1* 3'UTR in CM resistant vector) is digested with AscI and HindIII and KAWHIT03.0072 is digested with MluI and HindIII. The *FATB-1* 3'UTR is then directionally cloned into KAWHIT03.0072 to make KAWHIT03.0073 (soybean *FAD2-1A* intron, *FAD3-1A* 3'UTR, *FATB-1* 3'UTR with *FAD3-1A* intron #5).

25 Step 11 – KAWHIT03.0073 is digested with BstXI and Sall and the fragment containing *FAD2-1A* intron, *FAD3-1A* 3'UTR and *FATB-1* 3'UTR is gel purified. In a different tube KAWHIT03.0073 is digested with XhoI and Sse8387I. The intron/3'UTR fragment is then cloned back into KAWHIT03.0073 in the opposite orientation on the other site of soybean *FAD3-1A* intron #5 to create KAWHIT03.0074 (soybean *FAD2-1A* intron #1 sense, soybean *FAD3-1A* 3'UTR sense, soybean *FATB-1* 3'UTR sense, soybean, spliceable soybean *FAD3-1A* intron #5, soybean *FATB-1* 3'UTR anti-sense, soybean *FAD3-1A* 3'UTR anti-sense, soybean *FAD2-1A* intron #1 anti-sense).

Step 12 – To link the RNAi construct to the 7S alpha' promoter and the TML 3', KAWHIT03.0074 and pMON68527 (7Sa'/TML3' cassette) are digested with SacI and ligated together to make pMON68563 (7S alpha' promoter - *FAD2-1A* intron #1 sense, soybean *FAD3-1A* 3'UTR sense, soybean *FATB-1* 3'UTR sense, spliceable soybean *FATB-1* 3'UTR anti-sense, soybean *FAD3-1A* 3'UTR anti-sense, soybean *FAD2-1A* intron #1 anti-sense – TML3').

Step 13 – To introduce the assembled RNAi construct into pMON70682, pMON68563 and pMON70682 are digested with NotI and ligated together to form pMON68536 comprising a 7S alpha' promoter operably linked to the double-stranded-RNA-forming construct of *FAD2-1A* intron #1 sense, soybean *FAD3-1A* 3'UTR sense, soybean *FATB-1* 3'UTR sense, spliceable soybean *FAD3-1A* intron #5, soybean *FATB-1* 3'UTR anti-sense, soybean *FAD3-1A* 3'UTR anti-sense, soybean *FAD2-1A* intron #1 anti-sense and TML3' terminator).

Step 14 – pMON68536 is then digested with AscI and RsrII and pMON68529 (which contains the selectable marker CP4 fused to the FMV promoter and the RBCS 3' and the soybean *FAD2* promoter driving the delta 9 desaturase) is digested with SanDI and AscI. The RNAi portion of pMON68536 is then directionally cloned into pMON68529 to create pMON68537 (7S alpha' promoter operably linked to the double-stranded-RNA-forming construct of *FAD2-1A* intron #1 sense, soybean *FAD3-1A* 3'UTR sense, soybean *FATB-1* 3'UTR sense, spliceable soybean *FAD3-1A* intron #5, soybean *FATB-1* 3'UTR anti-sense, soybean *FAD3-1A* 3'UTR anti-sense, soybean *FAD2-1A* intron #1 anti-sense and TML3' terminator and soybean *FAD2* promoter driving the delta 9 desaturase).

Example 8

The following fifteen steps illustrate the construction of vector pMON68539 (Figure 22) designed for plant transformation to suppress *FAD2*, *FAD3*, and *FATB* genes and over-express delta-9 desaturase and the KASIV enzyme in soybean. In particular, the construct comprises a 7S alpha promoter operably linked to soybean sense-oriented intron and 3'UTRs, i.e., a *FAD2-1A* intron #1, a *FAD3-1A* 3'UTR, a *FATB-1* 3'UTR, a hairpin loop-forming spliceable intron, and a complementary series of soybean anti-sense-oriented intron and 3'UTR's, i.e., a *FATB-1*

3'UTR, a *FAD3-1A* 3'UTR and a *FAD2-1A* intron #1, the soybean *FAD2* promoter driving the delta-9 desaturase, and the Napin promoter driving KASIV.

Step1 – The soybean *FAD3-1A* intron #5, which serves as the spliceable intron portion of the RNAi construct, is PCR amplified using soybean genomic DNA as template, with the following primers:

5' primer = 19037 = ACTAGTATATTGAGCTCATATTCCACTGCAGTGGATATTG
TTTAAACATAGCTAGCATATTACGCGTATATTATAACAAGCTTATATTCCCGGGATATT
GTCGACATATTAGCGGTACATTTTATTGCTTATTCAC

3' primer = 19045 = ACTAGTATATTGAGCTCATATTCCTGCAGGATATTCTCGAG
ATATTCACGGTAGTAATCTCCAAGAACTGGTTTTGCTGCTTGTGTCTGCAGTGAATC.

These primers add cloning sites to the 5' and 3' ends. To 5' end: SpeI, SacI, BstXI, PmeI, NheI, MluI, HindIII, XmaI, SmaI, SalI. To 3' end: SpeI, SacI, Sse8387I, XhoI. The soybean *FAD3-1A* intron #5 PCR product is cloned into pCR2.1, resulting in KAWHIT03.0065.

KAWHIT03.0065 is then digested with SpeI and the ends are filled with Pfu polymerase and pMON68526 (empty CM resistant vector) is digested with HindIII and the ends are filled with Pfu polymerase. KAWHIT03.0065 and pMON68526 are ligated to create pMON68541. (soybean *FAD3-1A* intron #5 with multiple cloning sites in Amp resistant vector).

Step 2 – The soybean *FATB-1* 3'UTR is amplified with the following primers: 18662= TTTAATTACAATGAGAATGAGATTTACTGC (adding Bsp120I to the 5' end) and 18661= GGGCCCGATTTGAAATGGTTAACG. The PCR product is then ligated into pCR2.1 to make KAWHIT03.0036.

Step 3 – KAWHIT03.0036 is then digested with Bsp120I and EcoRI and then cloned into the KAWHIT03.0032 (empty CM resistant vector with a multiple cloning site) to make KAWHIT03.0037 (*FATB-1* 3'UTR in empty CM resistant vector).

Step 4 – The soybean *FAD3-1A* 3'UTR is amplified with the following primers: 18639= GGGCCCGTTTCAAACCTTTTGG (adding Bsp120I to the 5' end) and 18549= TGAAACTGACAATTCAA. The PCR product is then ligated into pCR2.1 to make KAWHIT03.0034.

Step 5 – KAWHIT03.0034 is digested with Bsp120I and EcoRI and then ligated into KAWHIT03.0032 (empty CM resistant vector with a multiple cloning site) to make KAWHIT03.0035 (*FAD3-1A* 3'UTR in empty CM resistant vector).

Step 6 – The soybean *FAD 2-1A* intron #1 is PCR amplified using soybean genomic DNA as template, with the following primers: 5' primer = 18663 = GGGCCCGGTAAATTAATTGTGC (Adding Bsp120I site to 5' end); and 3' primer = 18664 = CTGTGTCAAAGTATAACAAGTTCAG. The resulting PCR product is cloned into pCR 2.1 creating KAWHIT03.0038.

Step 7 – Soybean *FAD 2-1A* intron #1 PCR product in KAWHIT03.0038 is cloned into KAWHIT03.0032 (empty CM resistant vector with a multiple cloning site) using the restriction sites Bsp120I and EcoRI. The resulting plasmid is KAWHIT03.0039 (soybean *FAD 2-1A* intron #1 in empty CM resistant vector).

Step 8 – KAWHIT03.0039 is digested with AscI and HindIII and pMON68541 (*FAD3-1A* intron #5 RNAi AMP resistant base vector) is digested with MluI and HindIII. The soybean *FAD 2-1A* intron #1 is then directionally cloned into pMON68541 (*FAD3-1A* intron #5 in Amp resistant vector with multiple cloning sites) to generate KAWHIT03.0071 (soybean *FAD2-1A* intron #1 with soybean *FAD3-1A* intron #5).

Step 9 – KAWHIT03.0035 (*FAD3-1A* 3'UTR in CM resistant vector) is digested with AscI and HindIII and KAWHIT03.0071 (*FAD2-1A* intron and *FAD3-1A* intron #5 RNAi AMP resistant base vector) is digested with MluI and HindIII. The soybean *FAD 3-1A* 3'UTR is then directionally cloned into KAWHIT03.0071 to generate KAWHIT03.0072 (soybean *FAD2-1A* intron #1 and *FAD3-1A* 3'UTR with soybean *FAD3-1A* intron #5).

Step 10 – KAWHIT03.0037 (*FATB-1* 3'UTR in CM resistant vector) is digested with AscI and HindIII and KAWHIT03.0072 is digested with MluI and HindIII. The *FATB-1* 3'UTR is then directionally cloned into KAWHIT03.0072 to make KAWHIT03.0073 (soybean *FAD2-1A* intron, *FAD3-1A* 3'UTR, *FATB-1* 3'UTR with *FAD3-1A* intron #5).

Step 11 – KAWHIT03.0073 is digested with BstXI and Sall and the fragment containing *FAD2-1A* intron, *FAD3-1A* 3'UTR and *FATB-1* 3'UTR is gel purified. In a different tube KAWHIT03.0073 is digested with XhoI and Sse8387I. The Intron/3'UTR fragment is then

cloned back into KAWHIT03.0073 in the opposite orientation on the other site of soybean *FAD3-1A* intron #5 to create KAWHIT03.0074 (soybean *FAD2-1A* intron #1 sense, soybean *FAD3-1A* 3'UTR sense, soybean *FATB-1* 3'UTR sense, soybean, spliceable soybean *FAD3-1A* intron #5, soybean *FATB-1* 3'UTR anti-sense, soybean *FAD3-1A* 3'UTR anti-sense, soybean *FAD2-1A* intron #1 anti-sense).

Step 12 – To link the RNAi construct to the 7S alpha' promoter and the TML 3', KAWHIT03.0074 and pMON68527 (7Sa'/TML3' cassette) are digested with SacI and ligated together to make pMON68563 (7S alpha' promoter - *FAD2-1A* intron #1 sense, soybean *FAD3-1A* 3'UTR sense, soybean *FATB-1* 3'UTR sense, spliceable soybean soybean *FATB-1* 3'UTR anti-sense, soybean *FAD3-1A* 3'UTR anti-sense, soybean *FAD2-1A* intron #1 anti-sense – TML3').

Step 13 – To introduce the assembled RNAi construct into pMON70682, pMON68563 and pMON70682 are digested with NotI and ligated together to form pMON68536 comprising a 7S alpha' promoter operably linked to the double-stranded-RNA-forming construct of *FAD2-1A* intron #1 sense, soybean *FAD3-1A* 3'UTR sense, soybean *FATB-1* 3'UTR sense, spliceable soybean *FAD3-1A* intron #5, soybean *FATB-1* 3'UTR anti-sense, soybean *FAD3-1A* 3'UTR anti-sense, soybean *FAD2-1A* intron #1 anti-sense and TML3' terminator).

Step 14 – pMON68536 is then digested with AscI and RsrII and pMON68529 (which contains the selectable marker CP4 fused to the FMV promoter and the RBCS 3' and the soybean *FAD2* promoter driving the delta 9 desaturase) is digested with SanDI and AscI. The RNAi portion of pMON68536 is then directionally cloned into pMON68529 to create pMON68537 (7S alpha' promoter operably linked to the double-stranded-RNA-forming construct of *FAD2-1A* intron #1 sense, soybean *FAD3-1A* 3'UTR sense, soybean *FATB-1* 3'UTR sense, spliceable soybean *FAD3-1A* intron #5, soybean *FATB-1* 3'UTR anti-sense, soybean *FAD3-1A* 3'UTR anti-sense, soybean *FAD2-1A* intron #1 anti-sense and TML3' terminator and soybean *FAD2* promoter driving the delta 9 desaturase).

Step 15 – pMON68537 is then digested with SanDI and AscI and pMON70683 (Napin driving KasIV) is digested with AscI and RsrII. The Napin/KasIV fragment is directionally cloned into pMON68537 to create pMON68539 (7S alpha' promoter operably linked to the

double-stranded-RNA-forming construct of *FAD2-1A* intron #1 sense, soybean *FAD3-1A* 3'UTR sense, soybean *FATB-1* 3'UTR sense, spliceable soybean *FAD3-1A* intron #5, soybean *FATB-1* 3'UTR anti-sense, soybean *FAD3-1A* 3'UTR anti-sense, soybean *FAD2-1A* intron #1 anti-sense and TML3' terminator, soybean *FAD2* promoter driving the delta 9 desaturase and Napin promoter driving KasIV.

Example 9

This example illustrates plant transformation to produce soybean plants with suppressed genes.

A transformation vector pMON68537 as prepared in Example 7 is used to introduce an intron/3'UTR double-stranded RNA-forming construct into soybean for suppressing the Δ 12 desaturase, Δ 15 desaturase, and *FATB* genes. Vector pMON68537 also contains the delta-9 desaturase (*FAB2*) and the CP4 genes. The vector is stably introduced into soybean (Asgrow variety A4922) via *Agrobacterium tumefaciens* strain ABI (Martinell, U.S. Patent No. 6,384,301). The CP4 selectable marker allows transformed soybean plants to be identified by selection on media containing glyphosate herbicide.

Fatty acid compositions are analyzed from seed of soybean lines transformed with the intron/3'UTR RNAi expression constructs using gas chromatography. R₁ pooled seed and R₁ single seed oil compositions demonstrate that the mono- and polyunsaturated fatty acid compositions are altered in the oil of seeds from transgenic soybean lines as compared to that of the seed from non-transformed soybean, (See Table 7). For instance, *FAD2* suppression provides plants with increased amount of oleic acid ester compounds; *FAD3* suppression provides plants with decreased linolenic acid ester compounds; and *FATB* suppression provides plants with reduced saturated fatty ester compounds, e.g. palmitates and stearates. Selections can be made from such lines depending on the desired relative fatty acid composition. Fatty acid compositions are analyzed from seed of soybean lines transformed with constructs using gas chromatography.

Example 10

This example illustrates plant transformation to produce soybean plants with suppressed genes.

A transformation vector pMON68539 as prepared in Example 3 is used to introduce an intron/3'UTR double-stranded RNA-forming construct into soybean for suppressing the $\Delta 12$ desaturase, $\Delta 15$ desaturase, and *FATB* genes. Vector pMON68539 also contains the KasVI and the CP4 genes. The vector is stably introduced into soybean (Asgrow variety A4922) via *Agrobacterium tumefaciens* strain ABI (Martinell, U.S. Patent No. 6,384,301). The CP4 selectable marker allows transformed soybean plants to be identified by selection on media containing glyphosate herbicide.

Fatty acid compositions are analyzed from seed of soybean lines transformed with the intron/3'UTR RNAi expression constructs using gas chromatography. R₁ pooled seed and R₁ single seed oil compositions demonstrate that the mono- and polyunsaturated fatty acid compositions were altered in the oil of seeds from transgenic soybean lines as compared to that of the seed from non-transformed soybean (See Table 8). For example, *FAD2* suppression provides plants with increased oleic acid ester compounds; *FAD3* suppression provides plants with decreased linolenic acid ester compounds; and *FATB* suppression provides plants with reduced saturated fatty ester compounds, e.g. palmitates and stearates. Selections can be made from such lines depending on the desired relative fatty acid composition. Fatty acid compositions are analyzed from seed of soybean lines transformed with constructs using gas chromatography.

Table 7. Fatty acid composition of R1 single seeds from pMON68537 Events

Construct	Event	18:1	18:3	16:0	18:0	18:2
PMON68537	GM_A36305	74.92	4.42	6.35	2.93	10.24
PMON68537	GM_A36305	74.8	4.33	6.57	2.93	10.23
PMON68537	GM_A36305	74.43	3.95	5.98	2.82	11.81
PMON68537	GM_A36305	73.32	3.99	6.79	3.24	11.48
PMON68537	GM_A36305	72.87	4.33	7.06	3.08	11.7
PMON68537	GM_A36305	16.63	9.53	13.5	4.06	55.31
PMON68537	GM_A36305	16.52	9.61	13.92	4.24	54.79
PMON68537	GM_A36305	15.67	9.66	13.64	4.19	55.89
PMON68537	GM_A36306	77.45	3.93	6.76	2.47	8.4

Construct	Event	18:1	18:3	16:0	18:0	18:2
PMON68537	GM_A36306	74.51	4.38	6.58	2.47	10.94
PMON68537	GM_A36306	73.21	4.64	7.04	3.08	11.04
PMON68537	GM_A36306	72.78	4.4	6.97	2.55	12.21
PMON68537	GM_A36306	71.67	4.76	6.94	3.25	12.2
PMON68537	GM_A36306	71.01	4.86	7.64	3.05	12.41
PMON68537	GM_A36306	69.72	4.76	7.66	2.95	13.75
PMON68537	GM_A36306	17.41	8.88	13.35	3.85	55.63
PMON68537	GM_A36307	77.22	3.71	6.8	2.77	8.5
PMON68537	GM_A36307	76.79	3.65	6.76	2.85	8.75
PMON68537	GM_A36307	71.44	4.54	7.2	3.58	12.17
PMON68537	GM_A36307	18.83	8.62	13.94	4.02	53.61
PMON68537	GM_A36307	18.81	8.38	13.27	3.7	54.97
PMON68537	GM_A36307	15.68	9.97	14.06	4.55	54.79
PMON68537	GM_A36307	15.28	10.64	14.68	4.43	53.97
PMON68537	GM_A36307	14.08	9.36	14.39	4.31	56.89
PMON68537	GM_A36309	78.67	3.53	6.09	2.5	8.18
PMON68537	GM_A36309	75.43	3.96	6.7	2.53	10.3
PMON68537	GM_A36309	71.41	4.19	6.92	2.74	13.67
PMON68537	GM_A36309	70.51	4.14	6.85	3.16	14.33
PMON68537	GM_A36309	67.51	5.01	7.45	3.15	15.69
PMON68537	GM_A36309	66.99	4.92	7.15	3.9	15.79
PMON68537	GM_A36309	20.09	8.46	12.41	5	52.97
PMON68537	GM_A36309	15.15	9.73	14.61	3.85	55.79
PMON68537	GM_A36310	74.28	4.77	7.31	1.85	10.9
PMON68537	GM_A36310	74.03	5.43	8.23	1.63	9.66
PMON68537	GM_A36310	73.07	5.09	7.37	1.76	11.75
PMON68537	GM_A36310	71.83	5.04	7.78	1.86	12.54
PMON68537	GM_A36310	68.01	6.26	9.8	1.97	13.13
PMON68537	GM_A36310	67.22	6.28	8.71	3.28	13.45
PMON68537	GM_A36310	65.37	6.87	10.01	1.94	14.9
PMON68537	GM_A36310	15.76	10.09	13.4	4.28	55.52
PMON68537	GM_A36311	77.87	3.56	5.9	2.46	9.05
PMON68537	GM_A36311	75.8	3.87	5.91	2.93	10.22
PMON68537	GM_A36311	75.61	3.71	6.21	2.56	10.75
PMON68537	GM_A36311	73.68	4.06	6	3.09	11.98
PMON68537	GM_A36311	72.66	4.11	6.41	3.14	12.48
PMON68537	GM_A36311	70.89	4.39	6.52	3.11	13.93
PMON68537	GM_A36311	70.82	3.97	6.52	3.18	14.29
PMON68537	GM_A36311	16.67	9.39	13.65	4.44	54.77
PMON68537	GM_A36312	78.32	4.3	6.36	1.79	8.16
PMON68537	GM_A36312	77.55	4.46	6.51	2.13	8.23
PMON68537	GM_A36312	77.43	4.17	6.31	1.81	9.24
PMON68537	GM_A36312	76.98	4.29	6.25	2.27	9.05
PMON68537	GM_A36312	76.43	4.55	6.82	2.16	8.96
PMON68537	GM_A36312	76.38	4.5	6.46	2.04	9.54
PMON68537	GM_A36312	75.25	4.27	6.41	1.97	11.06
PMON68537	GM_A36312	18.24	9.43	13.6	3.07	54.75
PMON68537	GM_A36313	80.18	4.07	6.17	2.59	5.85
PMON68537	GM_A36313	79.96	4.16	6.03	2.59	6.11

Construct	Event	18:1	18:3	16:0	18:0	18:2
PMON68537	GM_A36313	78.88	3.9	5.6	2.8	7.65
PMON68537	GM_A36313	78.76	3.92	5.44	2.91	7.82
PMON68537	GM_A36313	77.64	4.22	5.88	2.9	8.25
PMON68537	GM_A36313	76.15	4.14	6.06	3.13	9.42
PMON68537	GM_A36313	19.05	8.87	13.45	3.71	54.03
PMON68537	GM_A36313	18.47	8.46	13.13	3.63	55.41
PMON68537	GM_A36314	80.27	3.17	5.77	3.4	6.03
PMON68537	GM_A36314	79.66	3.24	5.72	3.19	6.91
PMON68537	GM_A36314	79.5	3.45	5.83	3.23	6.74
PMON68537	GM_A36314	77.42	3.52	5.76	3.57	8.42
PMON68537	GM_A36314	77.33	3.71	6.36	3.34	8.01
PMON68537	GM_A36314	76.83	3.71	6.38	3.24	8.59
PMON68537	GM_A36314	16.6	9.3	12.63	4.43	55.99
PMON68537	GM_A36314	15.26	8.59	13.71	4.54	56.84
PMON68537	GM_A36315	20.21	8.25	13.61	3.59	53.37
PMON68537	GM_A36315	17.47	9.22	13.46	3.35	55.57
PMON68537	GM_A36315	16.75	9.3	13.61	3.66	55.75
PMON68537	GM_A36315	16.54	9.18	13.54	3.88	55.9
PMON68537	GM_A36315	16.06	10.07	13.44	4.01	55.42
PMON68537	GM_A36315	16.05	9.58	12.82	4.25	56.29
PMON68537	GM_A36315	15.95	10.42	13.12	3.63	55.91
PMON68537	GM_A36315	15.5	10.22	13.25	3.78	56.3
PMON68537	GM_A36316	79.61	3.56	5.79	2.94	6.87
PMON68537	GM_A36316	75.11	4.01	6.45	3.44	9.76
PMON68537	GM_A36316	75.07	4.25	6.74	3.09	9.64
PMON68537	GM_A36316	73.92	3.97	6.53	3.56	10.75
PMON68537	GM_A36316	17.26	9.59	13.1	4.26	54.78
PMON68537	GM_A36316	17.15	9.03	12.81	4.04	55.97
PMON68537	GM_A36316	16.62	9.2	13.15	3.99	56.03
PMON68537	GM_A36316	16.6	9.44	13.19	3.95	55.84
PMON68537	GM_A36317	18.96	7.55	13.2	3.75	55.51
PMON68537	GM_A36317	16.19	9.43	13.33	3.96	56.04
PMON68537	GM_A36317	16.05	9.1	14.02	3.94	55.91
PMON68537	GM_A36317	15.33	9.4	13.91	4.22	56.11
PMON68537	GM_A36317	15.28	9.2	13.87	4.27	56.36
PMON68537	GM_A36317	14.58	10.15	13.74	4.38	56.15
PMON68537	GM_A36317	13.95	9.47	13.98	4.76	56.79
PMON68537	GM_A36317	13.91	9.88	14.26	4.62	56.25
PMON68537	GM_A36318	78.82	3.64	5.7	2.77	7.87
PMON68537	GM_A36318	77.94	3.73	5.9	2.94	8.29
PMON68537	GM_A36318	75.18	4.11	6.08	3.48	9.95
PMON68537	GM_A36318	75.1	3.93	6.02	3.04	10.75
PMON68537	GM_A36318	75.01	4.22	6.57	3.29	9.72
PMON68537	GM_A36318	74.17	4.2	6.51	3.27	10.68
PMON68537	GM_A36318	73.47	4.27	6.7	3.22	11.16
PMON68537	GM_A36318	30.57	10.54	14.83	5.55	36.92
PMON68537	GM_A36319	80	3.65	5.83	2.31	7.02
PMON68537	GM_A36319	79.89	3.65	5.64	2.35	7.26
PMON68537	GM_A36319	79.4	3.59	5.73	1.76	8.46

Construct	Event	18:1	18:3	16:0	18:0	18:2
PMON68537	GM_A36319	78	3.87	6.11	2.35	8.5
PMON68537	GM_A36319	76.08	4.22	6.5	2.35	9.74
PMON68537	GM_A36319	75.56	3.89	6.41	1.78	11.3
PMON68537	GM_A36319	75.26	4.27	6.47	2.37	10.5
PMON68537	GM_A36319	75.16	4.1	6.48	2.49	10.66
PMON68537	GM_A36320	81.27	3.19	5.84	2.4	6.09
PMON68537	GM_A36320	80.21	3.27	5.18	2.44	7.76
PMON68537	GM_A36320	79.64	3.38	5.5	2.67	7.63
PMON68537	GM_A36320	79.46	3.38	5.82	2.67	7.42
PMON68537	GM_A36320	78.5	3.59	6.24	2.49	8
PMON68537	GM_A36320	73.83	3.79	6.72	2.78	11.74
PMON68537	GM_A36320	73.1	3.95	6.9	2.39	12.48
PMON68537	GM_A36320	22.99	8.03	12.19	4.81	50.89
PMON68537	GM_A36324	75.93	3.77	6.58	2.76	9.76
PMON68537	GM_A36324	75.1	4.05	7.01	2.83	9.8
PMON68537	GM_A36324	17.83	8.79	12.78	4.11	55.49
PMON68537	GM_A36324	16.46	8.88	12.84	4.48	56.29
PMON68537	GM_A36324	16.35	9.25	13.51	4.17	55.66
PMON68537	GM_A36324	15.25	8.99	13.73	4.28	56.69
PMON68537	GM_A36324	14.16	10.17	13.95	4.11	56.58
PMON68537	GM_A36324	13.59	9.87	14.61	4.5	56.33
PMON68537	GM_A36357	80.19	3.03	5.59	3.2	6.62
PMON68537	GM_A36357	79.78	3.19	5.51	3.24	6.89
PMON68537	GM_A36357	78.5	3.55	5.75	3.17	7.71
PMON68537	GM_A36357	77.48	3.68	5.71	3.55	8.23
PMON68537	GM_A36357	77.28	3.79	5.66	3.48	8.46
PMON68537	GM_A36357	77.1	3.51	5.43	3.65	8.99
PMON68537	GM_A36357	71.9	4.24	6.47	3.67	12.39
PMON68537	GM_A36357	17.66	9.32	13.26	4.21	54.51
PMON68537	GM_A36359	77.91	3.35	5.67	3.24	8.53
PMON68537	GM_A36359	77.85	3.29	5.42	3.29	8.87
PMON68537	GM_A36359	76.71	3.65	6.07	3.35	8.95
PMON68537	GM_A36359	71.73	4.01	6.79	3.49	12.68
PMON68537	GM_A36359	69.32	4.51	6.99	3.66	14.13
PMON68537	GM_A36359	68.63	4.44	6.91	3.76	14.89
PMON68537	GM_A36359	18.87	8.03	13.38	3.86	54.81
PMON68537	GM_A36359	16.81	9.83	13.08	4.68	54.55
PMON68537	GM_A36360	79.34	3.29	5.99	3.15	6.88
PMON68537	GM_A36360	75.42	3.47	6.47	3.08	10.26
PMON68537	GM_A36360	75.3	3.86	6.69	3.2	9.64
PMON68537	GM_A36360	74.51	3.8	6.39	3.32	10.67
PMON68537	GM_A36360	21.49	6.95	13.07	3.92	53.46
PMON68537	GM_A36360	20.05	7.4	13.09	3.83	54.57
PMON68537	GM_A36360	16.08	9.14	13.02	4.64	56.03
PMON68537	GM_A36360	15.86	9.07	13.44	4.49	56.04
PMON68537	GM_A36361	82.13	2.83	5.67	3.13	4.81
PMON68537	GM_A36361	80.99	3.2	5.79	3.01	5.64
PMON68537	GM_A36361	74.39	3.85	6.33	3.5	10.59
PMON68537	GM_A36361	18.01	8.46	13.18	3.92	55.41

Construct	Event	18:1	18:3	16:0	18:0	18:2
PMON68537	GM_A36361	17.99	8.11	13.05	4.09	55.7
PMON68537	GM_A36361	17.35	8.31	13.4	4	55.88
PMON68537	GM_A36361	16.81	10.2	12.9	4.32	54.87
PMON68537	GM_A36361	16.55	8.5	13.21	4.22	56.45
PMON68537	GM_A36362	78.05	3.89	6.29	2.81	7.76
PMON68537	GM_A36362	76.89	3.69	6.32	3.12	8.76
PMON68537	GM_A36362	76.1	4	6.57	3.02	9.24
PMON68537	GM_A36362	76.01	4.08	6.24	3.03	9.48
PMON68537	GM_A36362	75.86	3.76	5.68	3.56	9.95
PMON68537	GM_A36362	75.79	4.07	6.43	3.15	9.34
PMON68537	GM_A36362	74.89	4.14	6.63	3.11	10.07
PMON68537	GM_A36362	17.22	8.8	13.75	3.77	55.54
PMON68537	GM_A36363	79.15	3.57	6.2	3.03	6.84
PMON68537	GM_A36363	75.69	3.83	7.07	2.73	9.53
PMON68537	GM_A36363	73.97	4.22	6.82	3.39	10.33
PMON68537	GM_A36363	72.53	4.31	6.64	3.7	11.59
PMON68537	GM_A36363	68.42	4.5	7.05	3.95	14.79
PMON68537	GM_A36363	18.39	8.7	13.61	4.1	54.28
PMON68537	GM_A36363	17.54	8.87	14.08	4.07	54.56
PMON68537	GM_A36363	15.87	9.66	14.56	4.2	54.69
PMON68537	GM_A36365	78.79	3.11	5.87	1.27	9.9
PMON68537	GM_A36365	76.76	3.86	5.79	1.66	10.91
PMON68537	GM_A36365	75.41	3.49	6.06	1.83	12.15
PMON68537	GM_A36365	73.57	3.65	6.11	1.5	14.19
PMON68537	GM_A36365	71.55	3.56	6.62	1.24	16.08
PMON68537	GM_A36365	70.41	4	6.07	2.15	16.33
PMON68537	GM_A36365	66.66	3.9	6.84	1.5	20.21
PMON68537	GM_A36365	63.96	4.22	7.08	2.27	21.52
PMON68537	GM_A36366	75.44	4.33	6.49	3.21	9.32
PMON68537	GM_A36366	74.75	4.21	6.87	2.71	10.33
PMON68537	GM_A36366	74.69	4.65	6.91	3.06	9.65
PMON68537	GM_A36366	73.23	4.89	7.23	2.99	10.52
PMON68537	GM_A36366	72.53	4.76	7.42	3.26	10.85
PMON68537	GM_A36366	67.15	5.05	7.47	3.33	15.87
PMON68537	GM_A36366	65.81	5.6	7.9	3.37	16.09
PMON68537	GM_A36366	62.31	6.19	8.71	3.22	18.55
PMON68537	GM_A36367	80.56	3.3	6.07	2.58	6.34
PMON68537	GM_A36367	77.78	3.58	6.47	2.66	8.45
PMON68537	GM_A36367	77.78	3.46	6.25	2.84	8.51
PMON68537	GM_A36367	77.39	3.81	6.71	2.86	8.11
PMON68537	GM_A36367	77.32	3.74	6.17	3.12	8.47
PMON68537	GM_A36367	75.93	3.97	6.23	3.43	9.29
PMON68537	GM_A36367	72.82	4.09	6.85	3.25	11.88
PMON68537	GM_A36367	19.31	7.58	13.7	3.59	55
PMON68537	GM_A36410	21.67	7.62	13.38	3.43	53.1
PMON68537	GM_A36410	20.9	8.33	12.93	3.64	53.33
PMON68537	GM_A36410	20.21	8.04	13.28	3.86	53.66
PMON68537	GM_A36410	20.02	8.71	12.79	3.71	53.87
PMON68537	GM_A36410	18.96	8.95	13.3	3.77	54.15

Construct	Event	18:1	18:3	16:0	18:0	18:2
PMON68537	GM_A36410	18.18	8.98	13.56	3.74	54.66
PMON68537	GM_A36410	17.61	9.29	12.93	4.12	55.13
PMON68537	GM_A36410	16.78	9.8	13.78	3.92	54.83
PMON68537	GM_A36411	75.06	4.33	6.49	2.93	10.08
PMON68537	GM_A36411	74.32	4.46	6.76	2.96	10.38
PMON68537	GM_A36411	73.41	4.76	6.91	3.11	10.78
PMON68537	GM_A36411	73.24	4.87	7.28	2.89	10.67
PMON68537	GM_A36411	22.38	8.17	13.47	3.6	51.51
PMON68537	GM_A36411	18.26	9.07	14.14	3.81	54.02
PMON68537	GM_A36411	17.52	10.1	13.1	4.03	54.36
PMON68537	GM_A36411	17.02	9.71	13.45	4.02	54.89
A3244	A3244	18.29	7.79	13.69	4.15	55.08
A3244	A3244	17.54	8.19	13.32	4.32	55.57
A3244	A3244	17.13	8.13	13.21	4.46	56.04
A3244	A3244	15.47	9.56	13.04	4.43	56.46
A3244	A3244	15.17	8.95	13.79	4.3	56.78
A3244	A3244	15.05	9.03	14.16	4.01	56.8
A3244	A3244	13.51	10.07	12.95	5.07	57.3
A3244	A3244	13.49	9.91	13.31	4.56	57.67

Table 8. Fatty acid composition of R1 single seeds from pMON68539 Events

Construct	Event	16:0	18:0	18:1	18:2	18:3
PMON68539	GM_A36448	4.51	2.65	79.64	8.66	3.55
PMON68539	GM_A36448	4.62	2.64	78.35	9.99	3.77
PMON68539	GM_A36448	5.89	2.65	76.86	9.79	3.84
PMON68539	GM_A36448	4.92	2.62	72.61	14.61	4.01
PMON68539	GM_A36448	5.48	2.86	71.07	15.63	4.16
PMON68539	GM_A36448	13.5	4.2	16.28	56.86	8.29
PMON68539	GM_A36448	14.49	4.67	14.88	56.56	9.07
PMON68539	GM_A36449	5.16	2.42	81.91	6.54	3.12
PMON68539	GM_A36449	4.26	2.41	79.99	8.4	3.94
PMON68539	GM_A36449	4.26	2.72	79.07	9.32	3.38
PMON68539	GM_A36449	5.01	2.54	75.71	11.94	3.9
PMON68539	GM_A36449	4.34	2.76	75.07	12.75	4.16
PMON68539	GM_A36449	11.57	3.52	44.08	35.22	4.98
PMON68539	GM_A36449	13.42	3.84	21.35	52.38	8.17
PMON68539	GM_A36449	13.25	3.99	15.3	57.6	9.04
PMON68539	GM_A36450	3.28	2.6	82.21	7.26	3.95
PMON68539	GM_A36450	4.16	2.51	80.93	7.72	3.76
PMON68539	GM_A36450	4.3	3.42	78.78	8.43	4.22
PMON68539	GM_A36450	4.84	3.16	77.07	9.6	4.22
PMON68539	GM_A36450	5.11	3.1	75.21	10.98	4.49
PMON68539	GM_A36450	13.74	4.26	17.31	54.32	10.11
PMON68539	GM_A36450	13.82	4.34	17.13	54.96	9.47
PMON68539	GM_A36450	13.56	3.83	17.06	56.7	8.6
PMON68539	GM_A36705	9.73	1.83	75.04	8.23	4.27
PMON68539	GM_A36705	10.85	1.74	72.89	9.29	4.53

Construct	Event	16:0	18:0	18:1	18:2	18:3
PMON68539	GM_A36705	10.05	1.78	72.68	9.83	4.48
PMON68539	GM_A36705	10.02	1.77	72.57	10.04	4.36
PMON68539	GM_A36705	10.75	1.75	72.37	9.68	4.77
PMON68539	GM_A36705	10.58	1.78	70.35	11.64	4.43
PMON68539	GM_A36705	7.69	5.63	16.21	60.39	8.85
PMON68539	GM_A36705	8.02	5.69	15.58	60.65	8.86
	A3244	13.03	4.31	21.23	52.61	7.77
	A3244	12.69	3.98	20.71	55.12	6.53
	A3244	15.2	5.02	19.83	49.96	8.83
	A3244	12.63	4.84	19.55	53.18	8.66
	A3244	13.27	4.48	18.28	54.4	8.5
	A3244	13.22	4.91	17.38	54.73	8.63
	A3244	13.44	4.81	15.46	56.49	8.91